



# Rapid Soil Analysis by Mid-Infrared Spectroscopy Training Course

## 01. COURSE INTRODUCTION

Anthony Ringrose-Voase

May 2018

Agriculture and Food  
[www.csiro.au](http://www.csiro.au)



Australian Government  
Australian Centre for  
International Agricultural Research



# Training course objectives

To train participants in the use of mid-infrared spectroscopy (MIR) to rapidly characterize soil

- Role of rapid soil analysis in a soil survey context
- Operation of MIR
- Calibration of MIR and prediction of soil properties

# Course outline

## Seminar on “Rapid Soil Analysis by Mid-Infrared Spectroscopy”

1. What is MIR spectroscopy?
  - Demonstration of MIR operation (video)
2. MIR calibration
  - Demonstration of MIR calibration
  - Demonstration of soil property estimation by MIR
3. Role of rapid soil analysis in digital soil mapping
4. MIR calibration strategies

# Course outline

## Hands-on training course

1. Course outline Monday PM
2. What is MIR spectroscopy?
3. Role of rapid soil analysis in digital soil mapping
  
4. Setting up MIR spectrometer and software Tuesday AM
5. Sample preparation Tuesday PM
6. Safety
7. Acquiring MIR spectra from soil specimens
  
8. Calibration Wednesday
9. MIR calibration strategies
  
10. Calibration and optimization Thursday
  
11. Soil property estimation Friday



# Course outline

The course will consist of:

- Lectures
  - Background to theory and practice of MIR spectroscopy
  - Seminar
- Instructions
  - Demonstrations of practical aspects of MIR spectroscopy
- Practicals
  - Opportunities for participants to learn-by-doing

# Training course objectives

At the end of this course you will be able to:

- Understand the principles of MIR spectroscopy as applied to rapid soil analysis
- Be able to:
  - Prepare soil specimens for MIR spectroscopy
  - Acquire MIR spectra from soil specimens
  - Develop MIR calibrations for different soil properties
  - Predict soil properties from MIR spectra
- Understand the role of MIR spectroscopy in soil survey and land evaluation



# Rapid Soil Analysis by Mid-Infrared Spectroscopy Training Course

## 02. WHAT IS MIR SPECTROSCOPY?

Anthony Ringrose-Voase  
May 2018

Agriculture and Food  
[www.csiro.au](http://www.csiro.au)

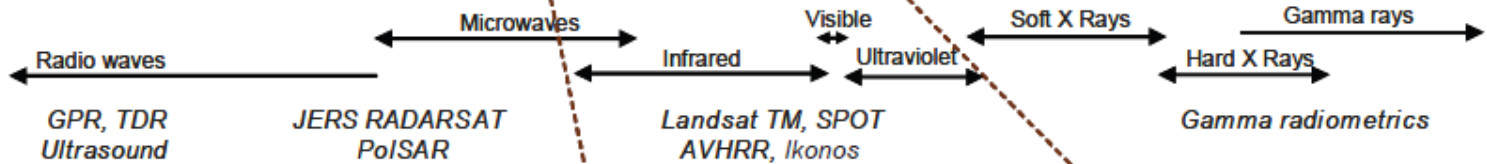
# Range of electromagnetic radiation available for rapid soil measurement

Frequency (Hz)

$10^3$   $10^4$   $10^5$   $10^6$   $10^7$   $10^8$   $10^9$   $10^{10}$   $10^{11}$   $10^{12}$   $10^{13}$   $10^{14}$   $10^{15}$   $10^{16}$   $10^{17}$   $10^{18}$   $10^{19}$   $10^{20}$   $10^{21}$   $10^{22}$   $10^{23}$

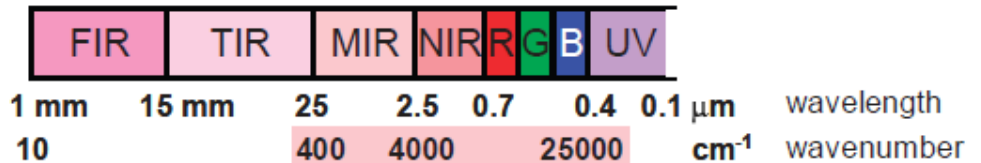
Wavelength (m)

$10^6$   $10^5$   $10^4$   $10^3$   $10^2$   $10$   $1$   $10^{-1}$   $10^{-2}$   $10^{-3}$   $10^{-4}$   $10^{-5}$   $10^{-6}$   $10^{-7}$   $10^{-8}$   $10^{-9}$   $10^{-10}$   $10^{-11}$   $10^{-12}$   $10^{-13}$   $10^{-14}$



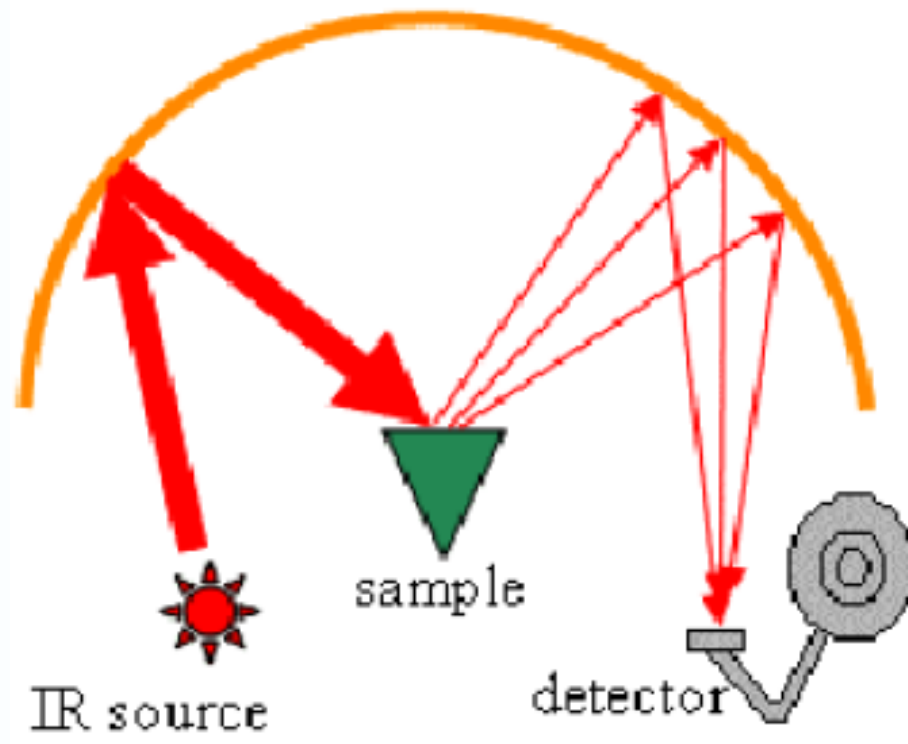
Energy of one photon (eV)

$10^{-11}$   $10^{-10}$   $10^{-9}$   $10^{-8}$   $10^{-7}$   $10^{-6}$   $10^{-5}$   $10^{-4}$   $10^{-3}$   $10^{-2}$   $10^{-1}$   $1$   $10$   $10^2$   $10^3$   $10^4$   $10^5$   $10^6$   $10^7$   $10^8$



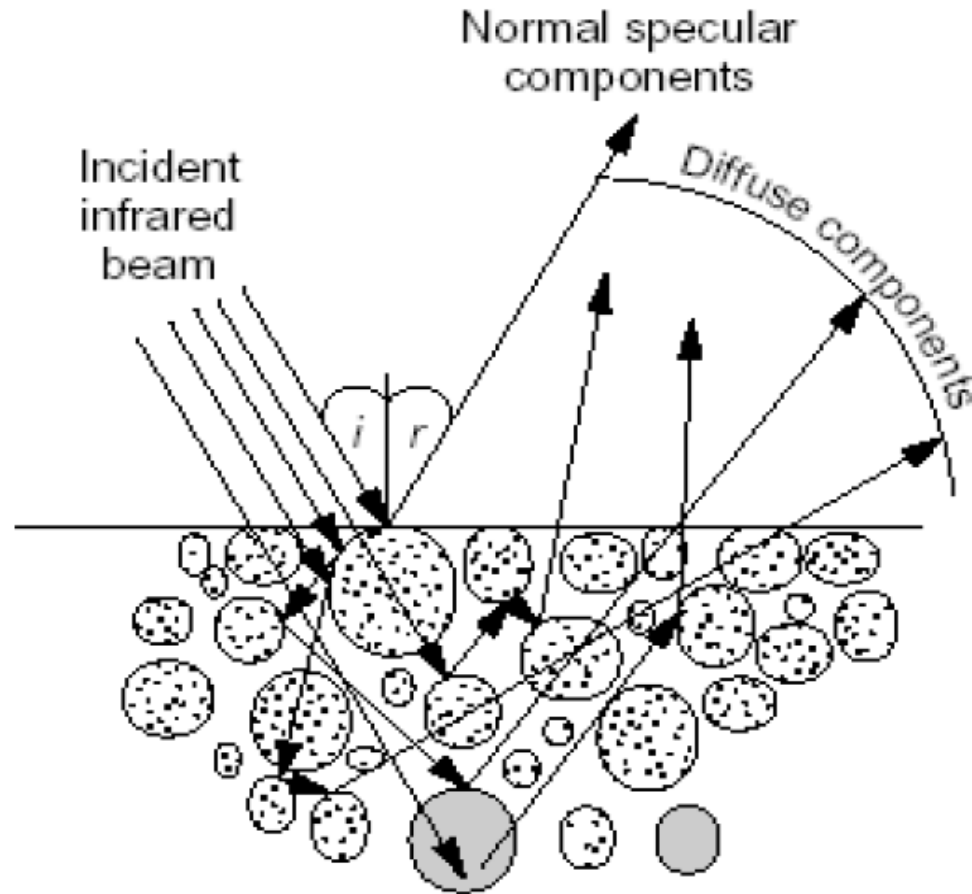
# MIR reflectance spectroscopy

- Infrared light is shone at a finely ground soil specimen
- Infrared light is scattered and reflected by soil
- Reflected infrared light is measured by a detector



# MIR reflectance spectroscopy

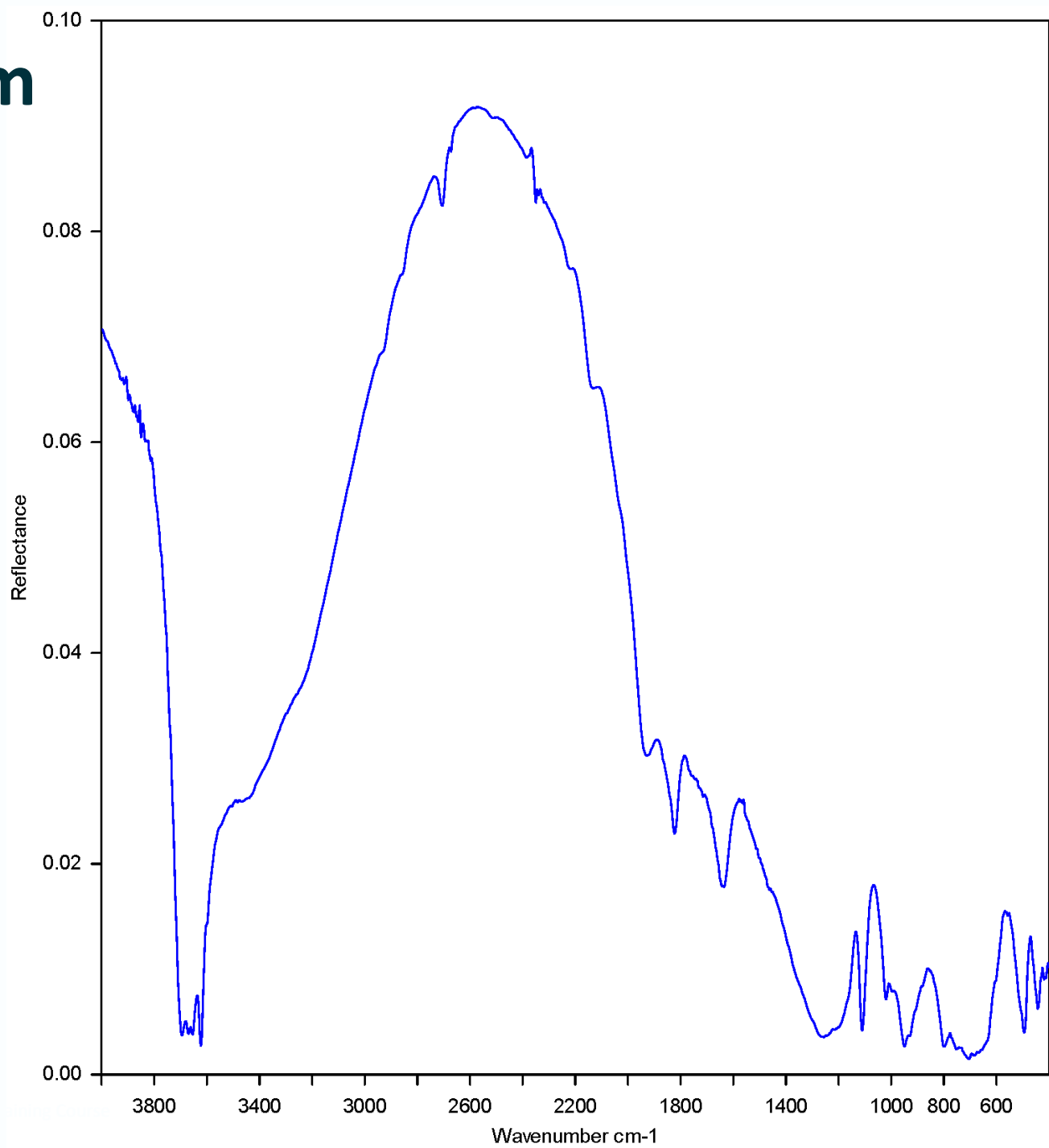
Needs dry fine soil particles



# MIR spectrum

Reflectance

— Kaolinite





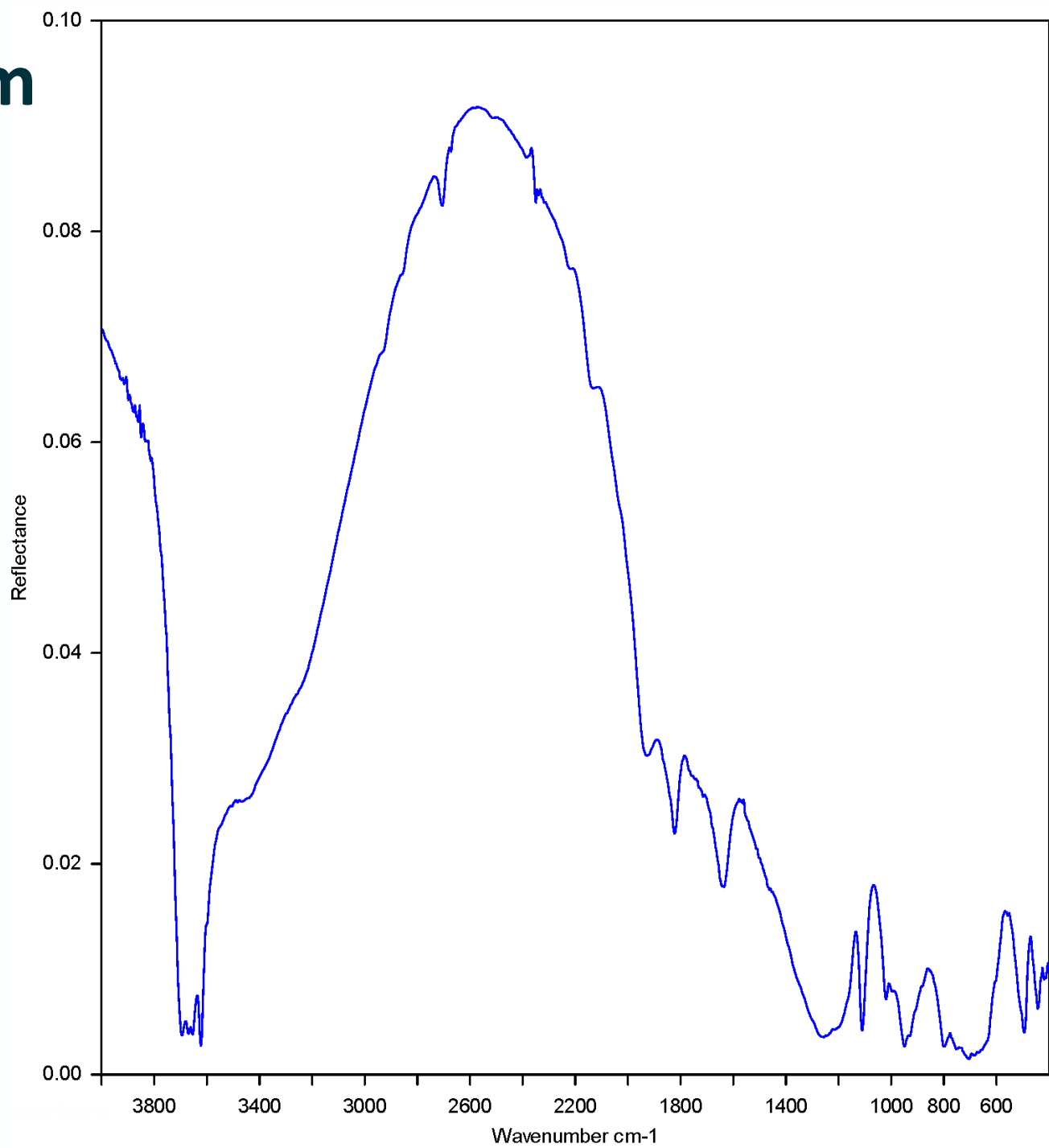
# MIR reflectance spectroscopy

- Some of the infrared light is absorbed by substances in the soil specimen
  - Different substances absorb specific infrared wavelengths
  - Which wavelengths are absorbed depends on chemical bonds of the substance
  - In the infrared light reflected by the specimen, the intensity of these wavelengths is reduced
- Reduction in intensity is proportional to the concentration of the substance
- The reflectance spectrum carries information about the concentrations of the substances in the soil specimen
  - Like a fingerprint

# MIR spectrum

Reflectance

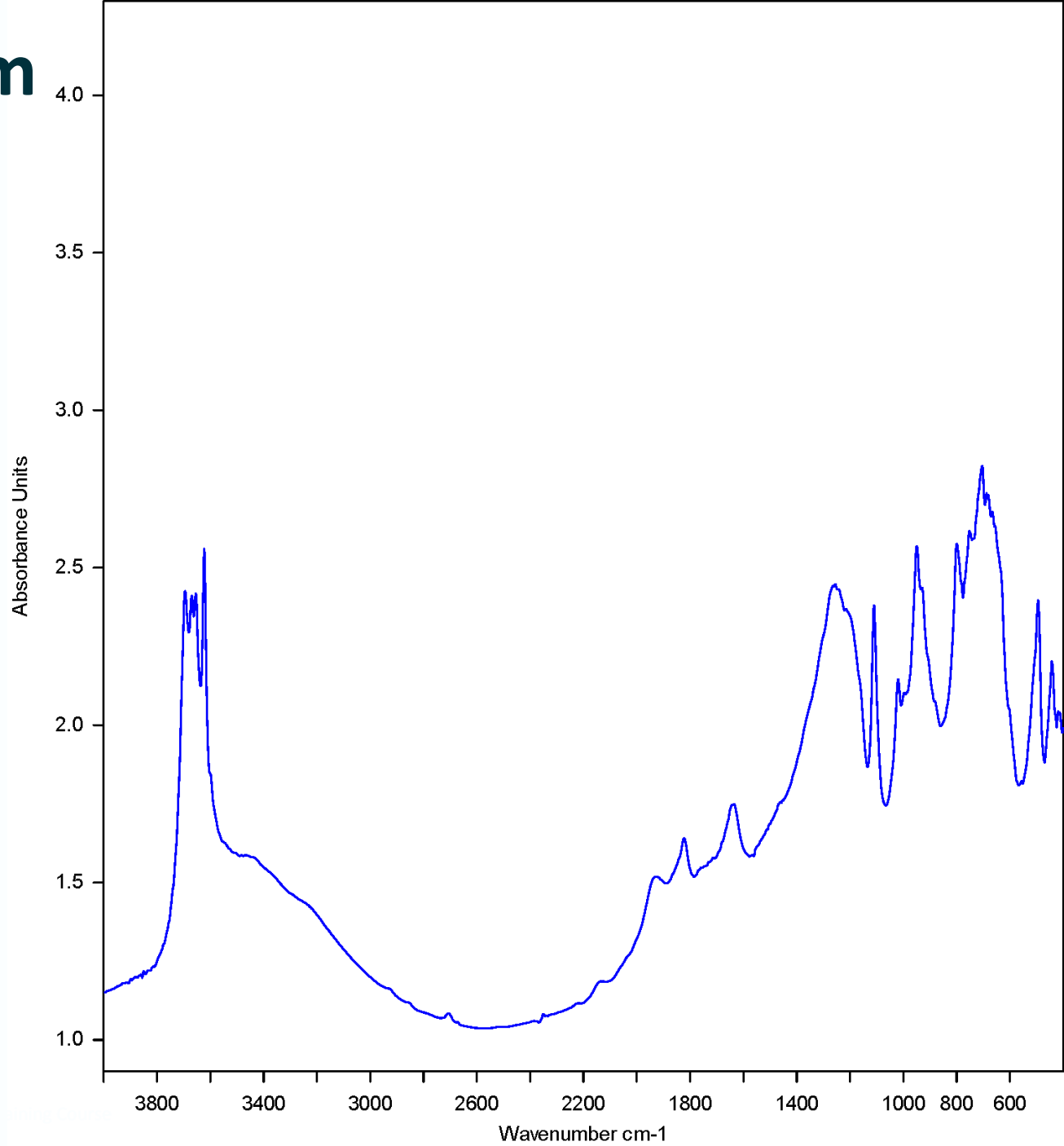
— Kaolinite



# MIR spectrum

Absorbance

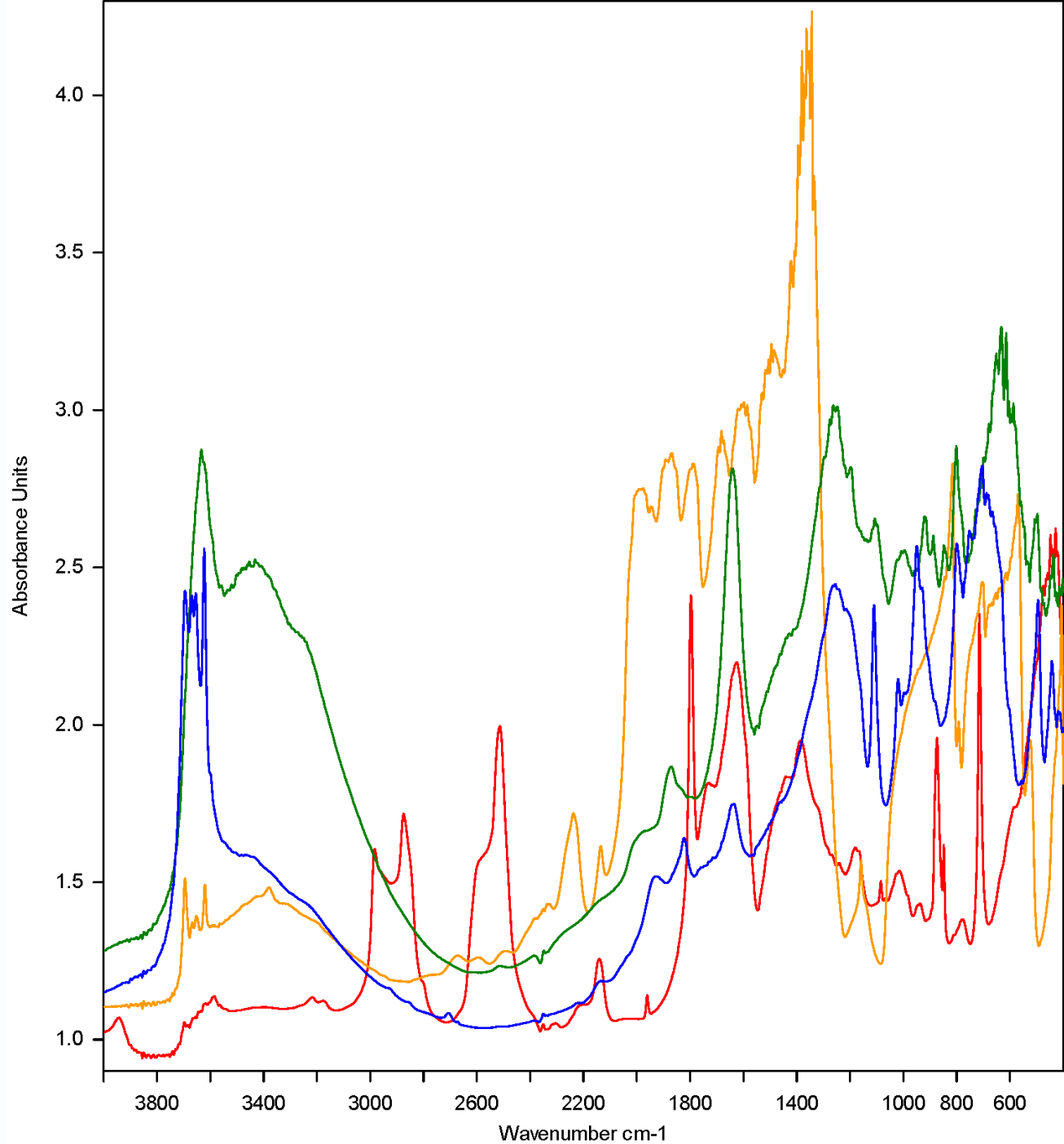
— Kaolinite



# MIR spectra





## Absorbance

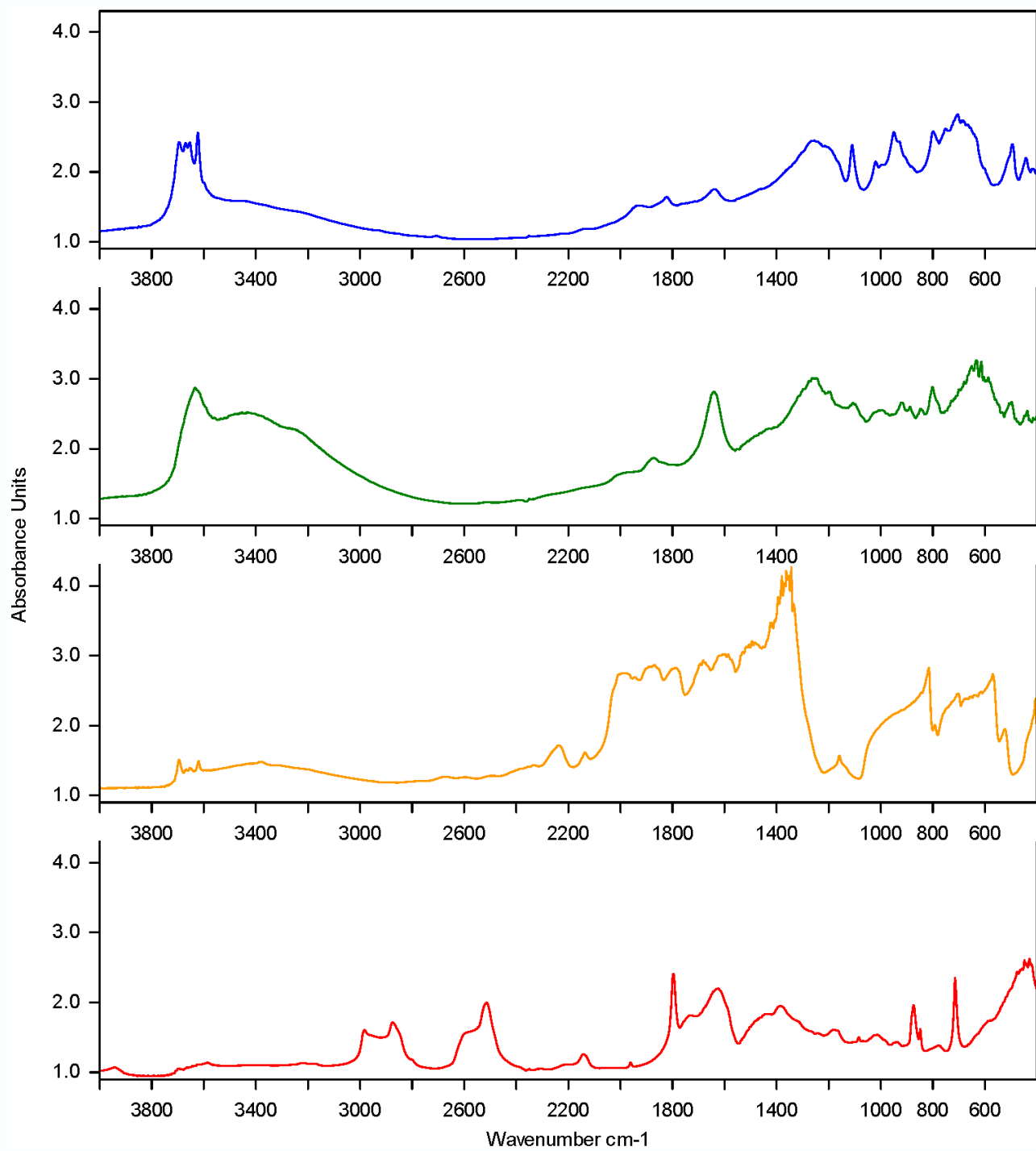
- Kaolinite
- Smectite
- Sand
- Carbonate



# MIR spectra

## Absorbance

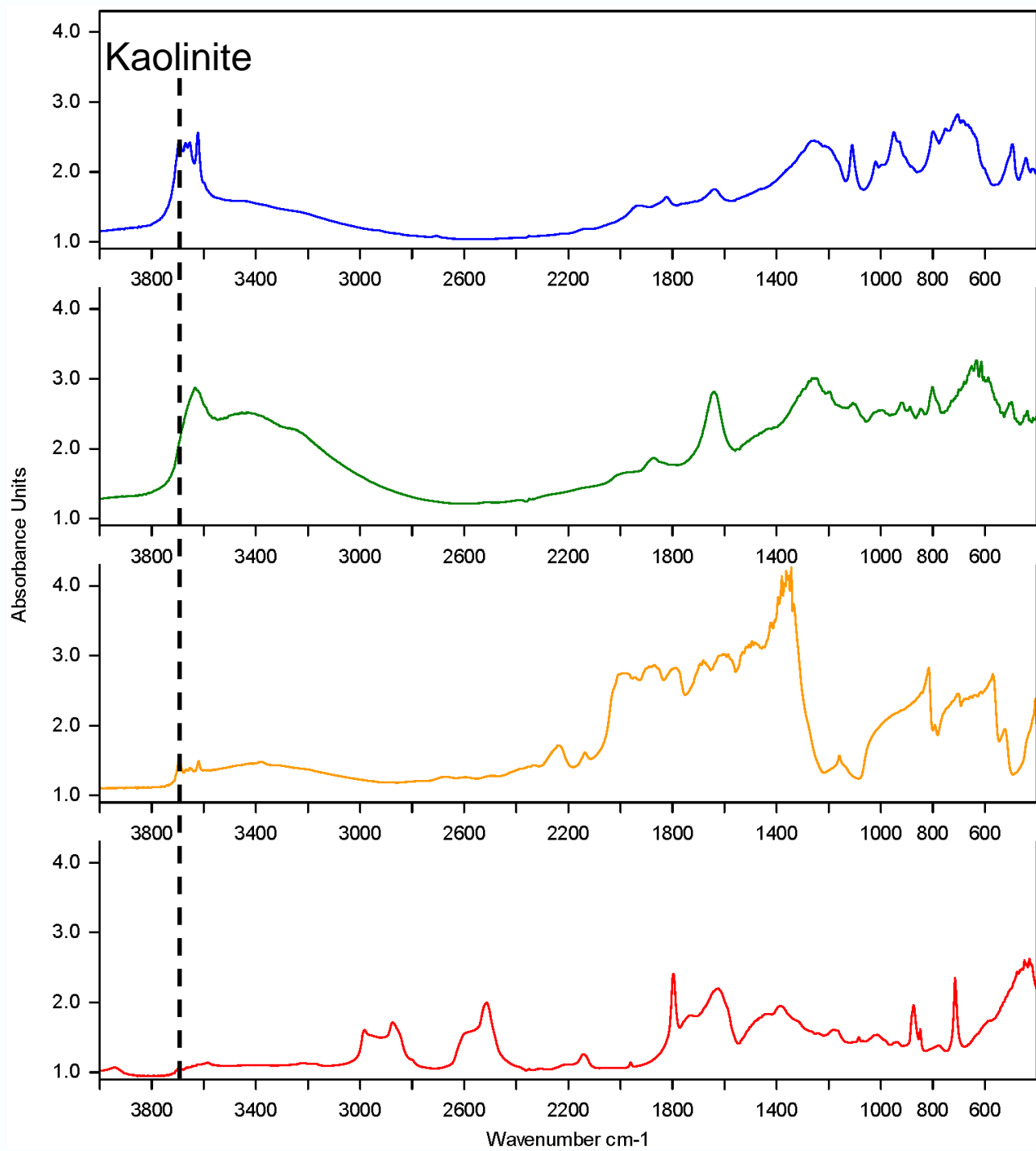
-  Kaolinite
-  Smectite
-  Sand
-  Carbonate



# MIR spectra

## Absorbance

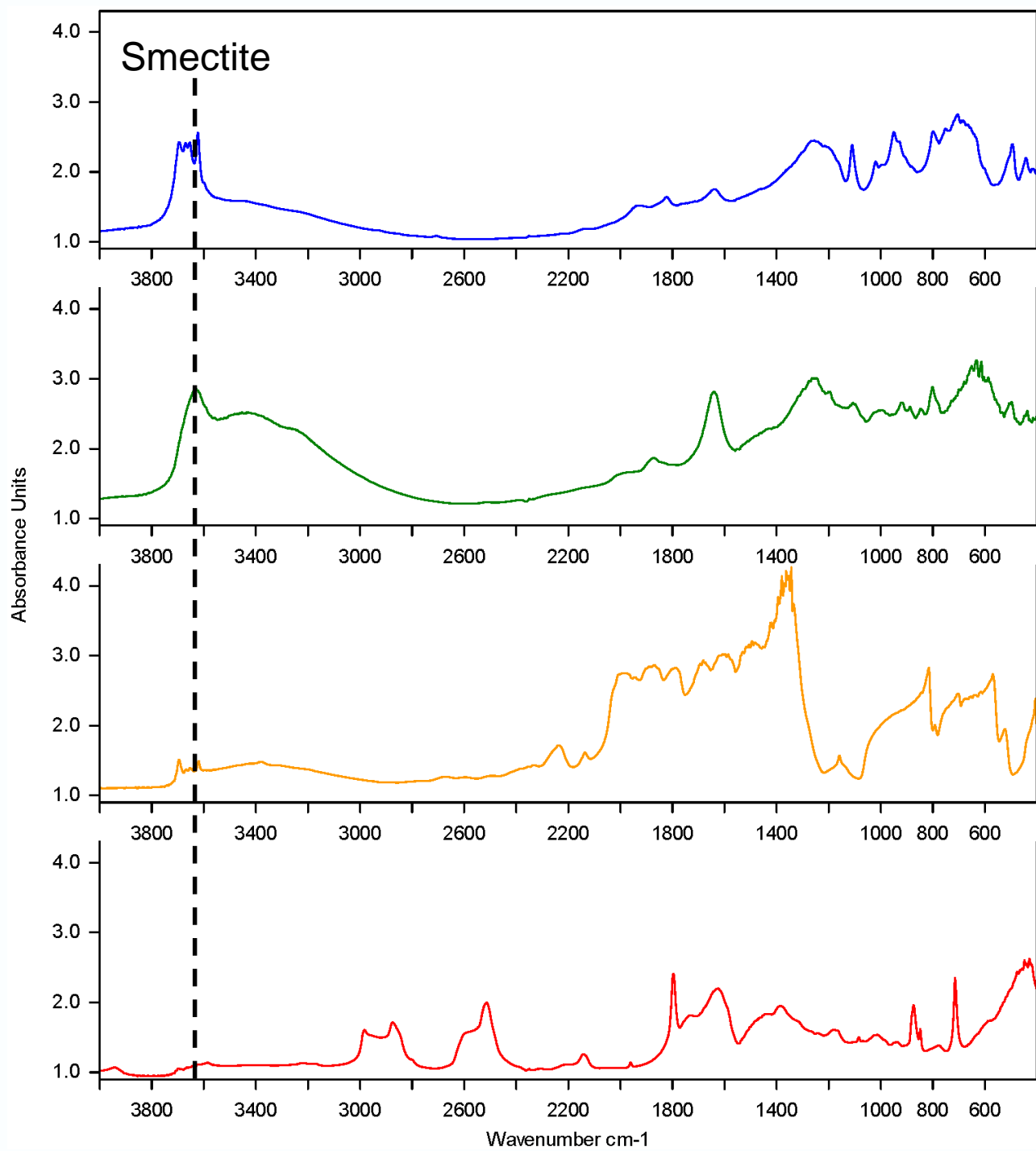
- Kaolinite
- Smectite
- Sand
- Carbonate



# MIR spectra

## Absorbance

- Kaolinite
- Smectite
- Sand
- Carbonate

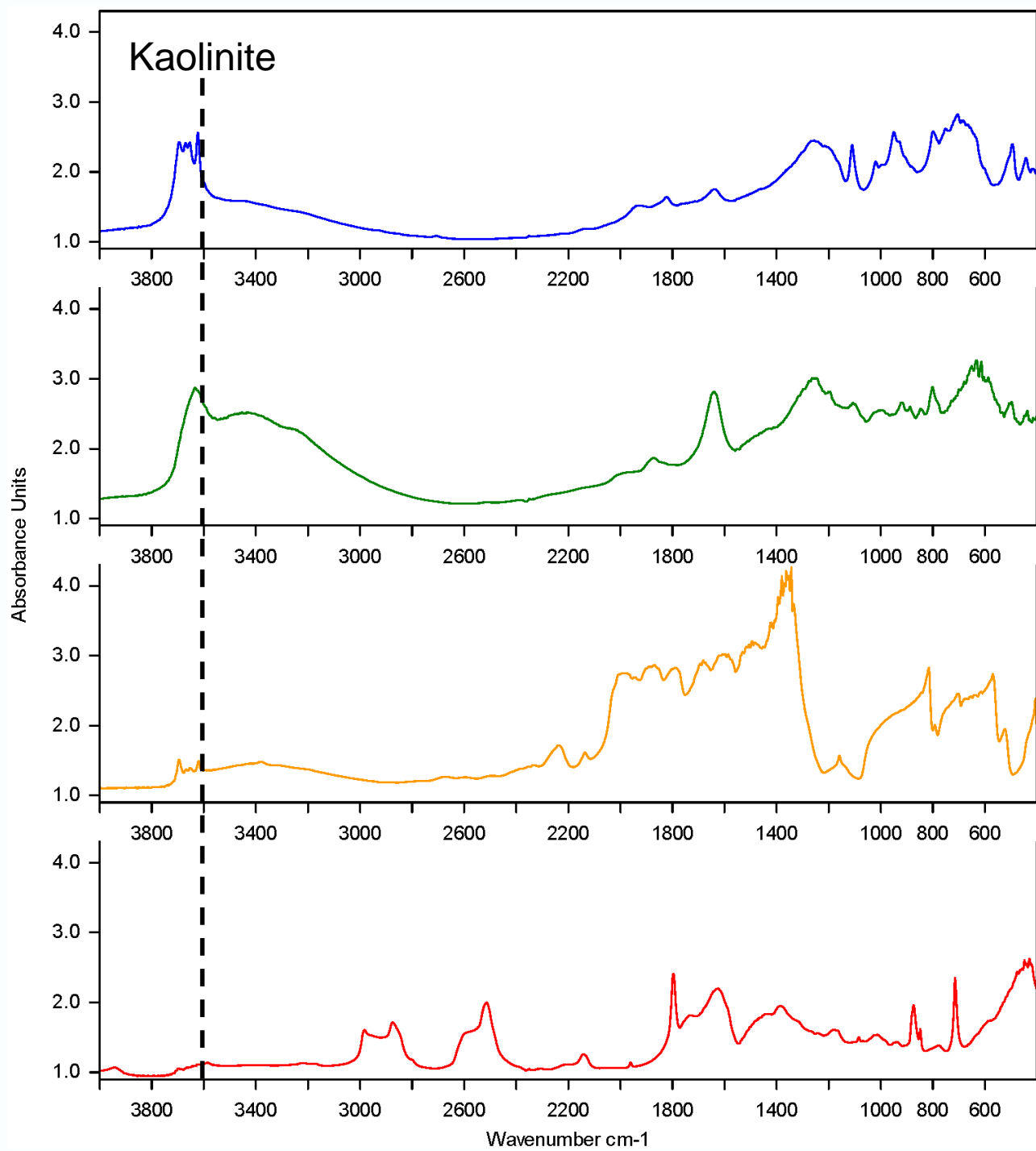




# MIR spectra

## Absorbance

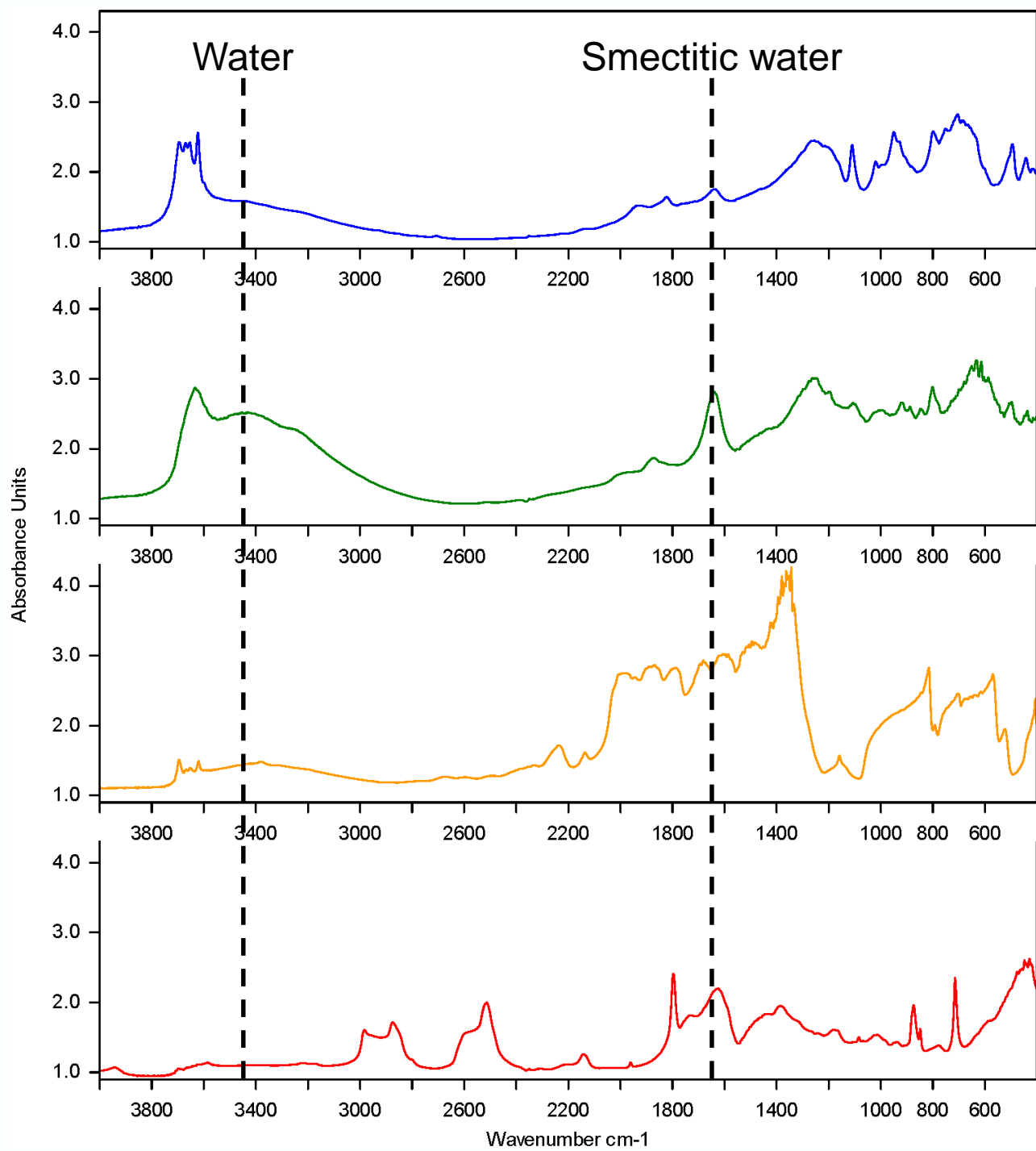
- Kaolinite
- Smectite
- Sand
- Carbonate



# MIR spectra

## Absorbance

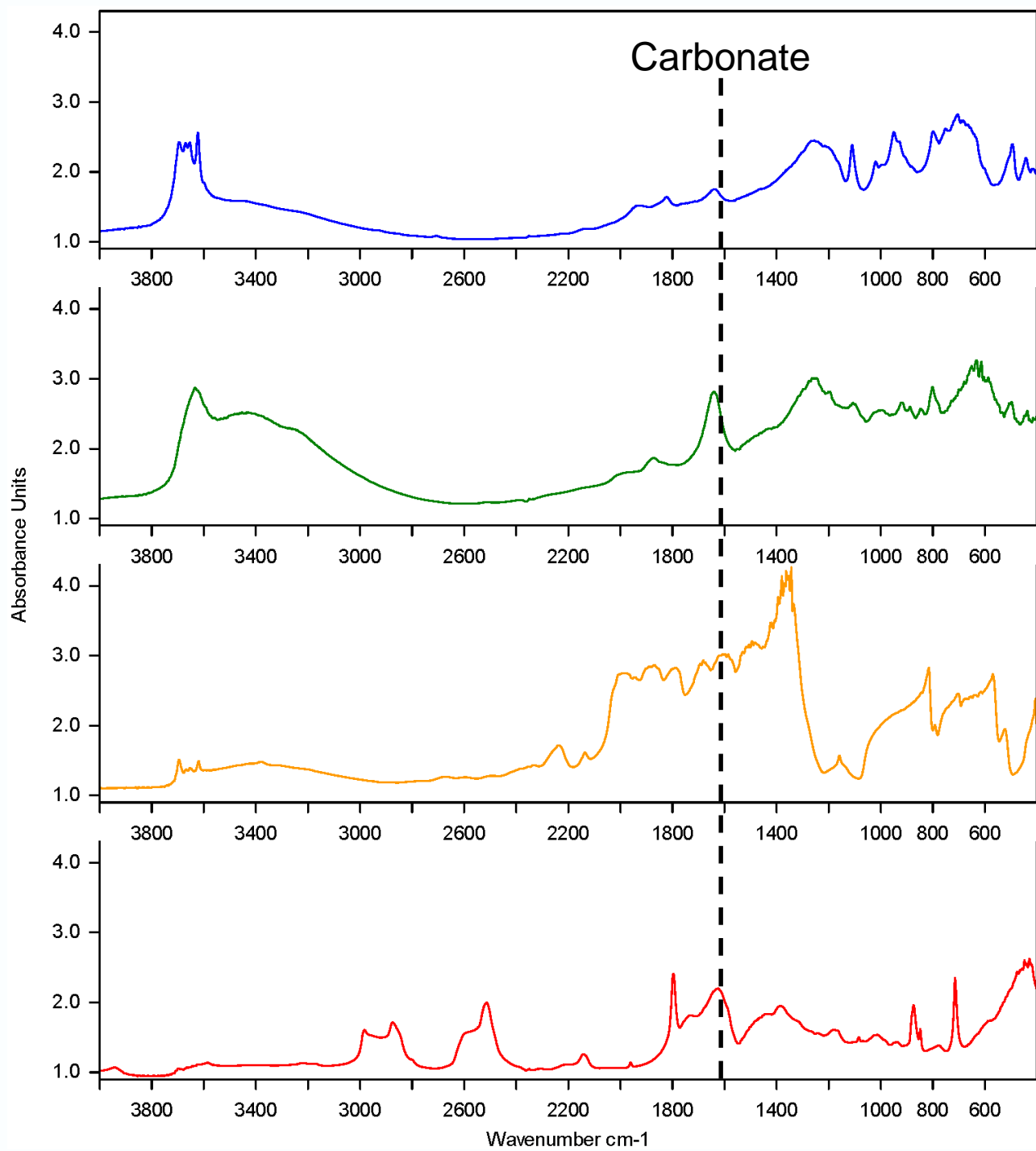
- Kaolinite
- Smectite
- Sand
- Carbonate



# MIR spectra





## Absorbance

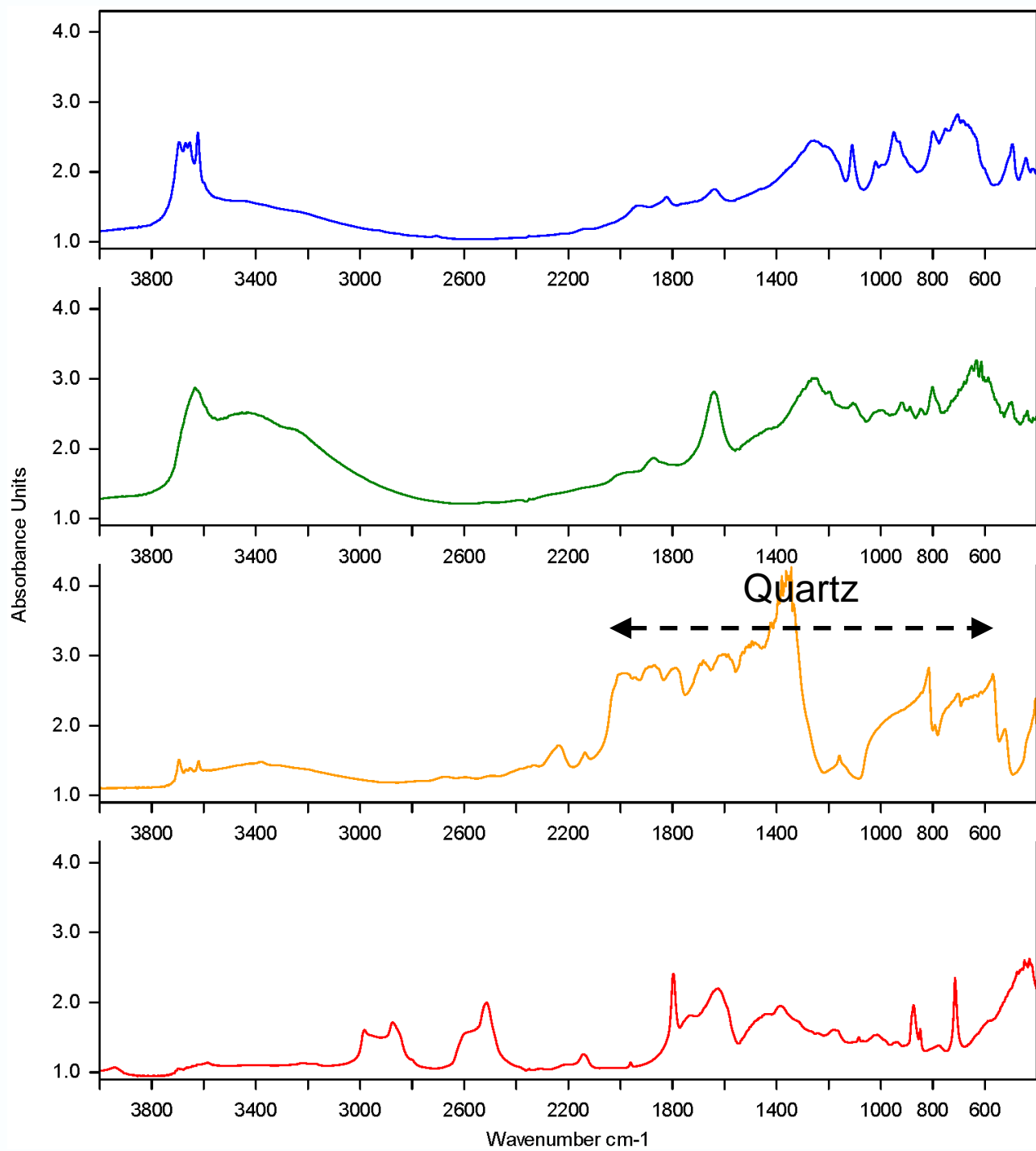
- Kaolinite
- Smectite
- Sand
- Carbonate



# MIR spectra

## Absorbance

-  Kaolinite
-  Smectite
-  Sand
-  Carbonate



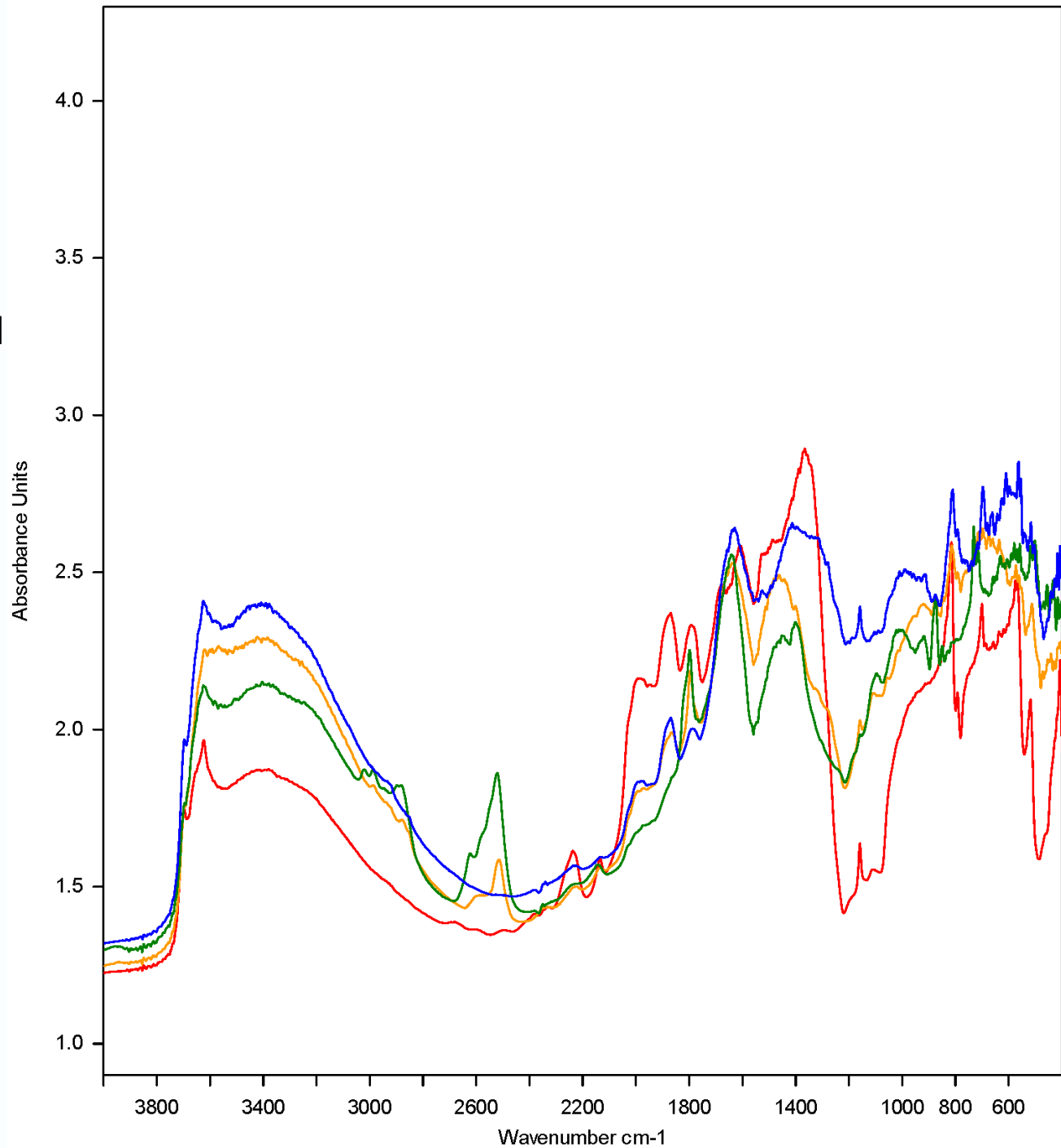
# MIR reflectance spectroscopy

- Spectra of real soil is a complicated mixture of peaks
  - Dependent on the concentrations of different substances

# MIR spectra

## Absorbance

- Clay topsoil
- Clay loam subsoil
- Silty loam topsoil
- Loamy sand topsoil



# MIR reflectance spectroscopy

- Spectra of real soil is a complicated mixture of peaks
  - Dependent on the concentrations of different substances
- These concentrations are usually estimated statistically
  - Process is called calibration
  - Uses a training set to ‘teach’ the system
    - Usually a small subset is selected
    - Training set is analysed by conventional laboratory methods
  - A statistical calibration is developed using “partial least squares”
    - Calibrations are developed for each soil property of interest
- This allows a range of soil properties can be estimated from the MIR spectra of the remaining soils



# MIR reflectance spectroscopy

- Substances that can be estimated include:
  - Clay
  - Silt
  - Sand
  - Organic carbon
  - Total nitrogen
  - pH
  - Exchangeable cations and CEC
  - Iron
- Other properties related to the above **may** be estimated in a given region
  - Phosphorus buffering index (PBI)
  - Plant available water capacity

# MIR reflectance spectroscopy

## Benefits

- Minimum soil preparation
  - Only drying and grinding to  $<0.5$  mm
  - No extracts or wet chemistry
- Requires only small sample (teaspoon)
  
- MIR spectrum acquired for each soil sample is used to estimate many soil properties
  - Each soil property requires a one-off calibration against conventional measurements
  - Once calibrations are developed several soil properties can be predicted simultaneously
  
- Stored spectra can be used to predict additional soil properties when new calibrations become available

# MIR reflectance spectroscopy

## Benefits

- Spectrum for one sample acquired in < 5 minutes
  - Allows rapid estimation of many soil properties for large number of samples
- For example: Soil survey of Pyawbwe Township, Myanmar produced 4000 samples
  - Characterisation by conventional analysis would not be possible
    - Time
    - Cost
  - Characterisation by MIR spectroscopy
    - + conventional analysis of subset about 100 samples
  - Cost conventional analysis: AUD 170 per sample
    - Total AUD 17,000
  - If used to calibrate MIR and estimate properties for remaining 3900 samples
    - Effective cost:  $\text{AUD } 17,000 / 4000 = \text{AUD } 4.25$
- Cost of MIR spectrometer is less than many conventional analytical instruments

# Deployment of MIR capability

Recent technological advances mean MIR instruments are relatively inexpensive to buy and run

- Will be commissioned:
  - Koronivia Research Station
- Equipment:
  - Small oven to air-dry samples at 60°C
  - Several agate mortars & pestles
  - Small MIR spectrometer (size of a shoe box)
  - Computer to
    - control spectrometer
    - capture results
    - develop calibrations





# Rapid Soil Analysis by Mid-Infrared Spectroscopy Training Course

## 03. ROLE OF RAPID SOIL ANALYSIS IN DIGITAL SOIL MAPPING

Anthony Ringrose-Voase

May 2018

Agriculture and Food

[www.csiro.au](http://www.csiro.au)



Australian Government

Australian Centre for  
International Agricultural Research



# Types of soil measurement

In **conventional** soil survey the information collected at each observation site includes:

- Soil profile morphology descriptions: All sites
- Laboratory measurements of soil properties: A few 'representative' sites
  - Laboratory analysis is: time-consuming  
expensive

# Laboratory analysis in a survey context

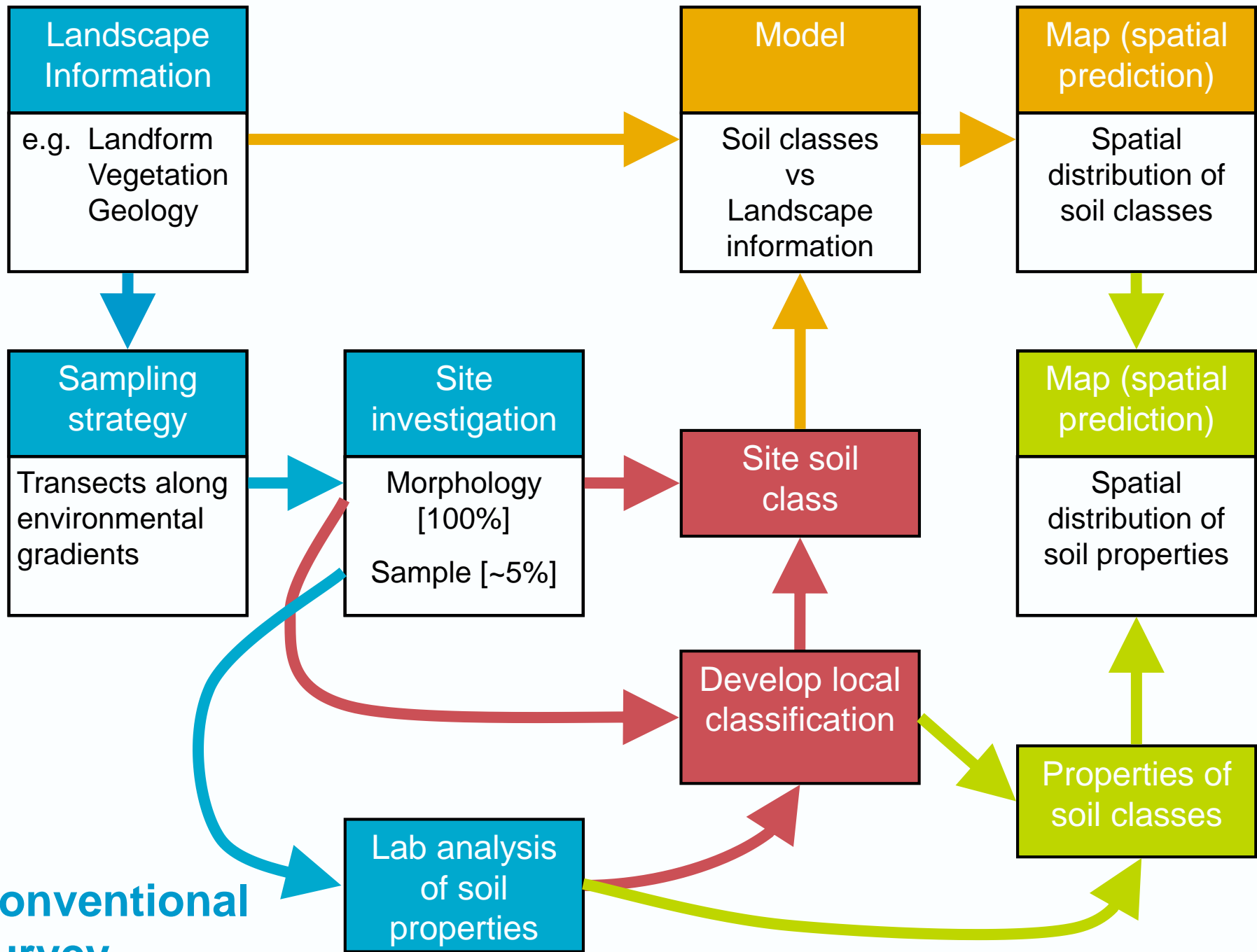
## Uses of laboratory analysis

- Assisting with soil classification
  - Required by some taxonomic systems (including Soil Taxonomy)
- Determine properties of each soil class

## Why are they necessary?

- Because they directly reflect the function of the soil
  - Usually more important than morphology
- Required for some aspects of land suitability assessment
- Required to guide advice on land management practices





# Types of soil measurement

In **conventional** soil survey the information collected at each observation site includes:

- Soil profile morphology descriptions: All sites
- Laboratory measurements of soil properties: A few 'representative' sites

In **digital** soil surveys the collected information collected:

- Soil profile morphology descriptions: All sites
- **Rapid soil measurements:** All sites
- Laboratory measurements of soil properties: A few 'calibration' specimens

# Role of rapid measurement

Measurement is 'rapid'

- Like morphology, can be done on all samples from all sites

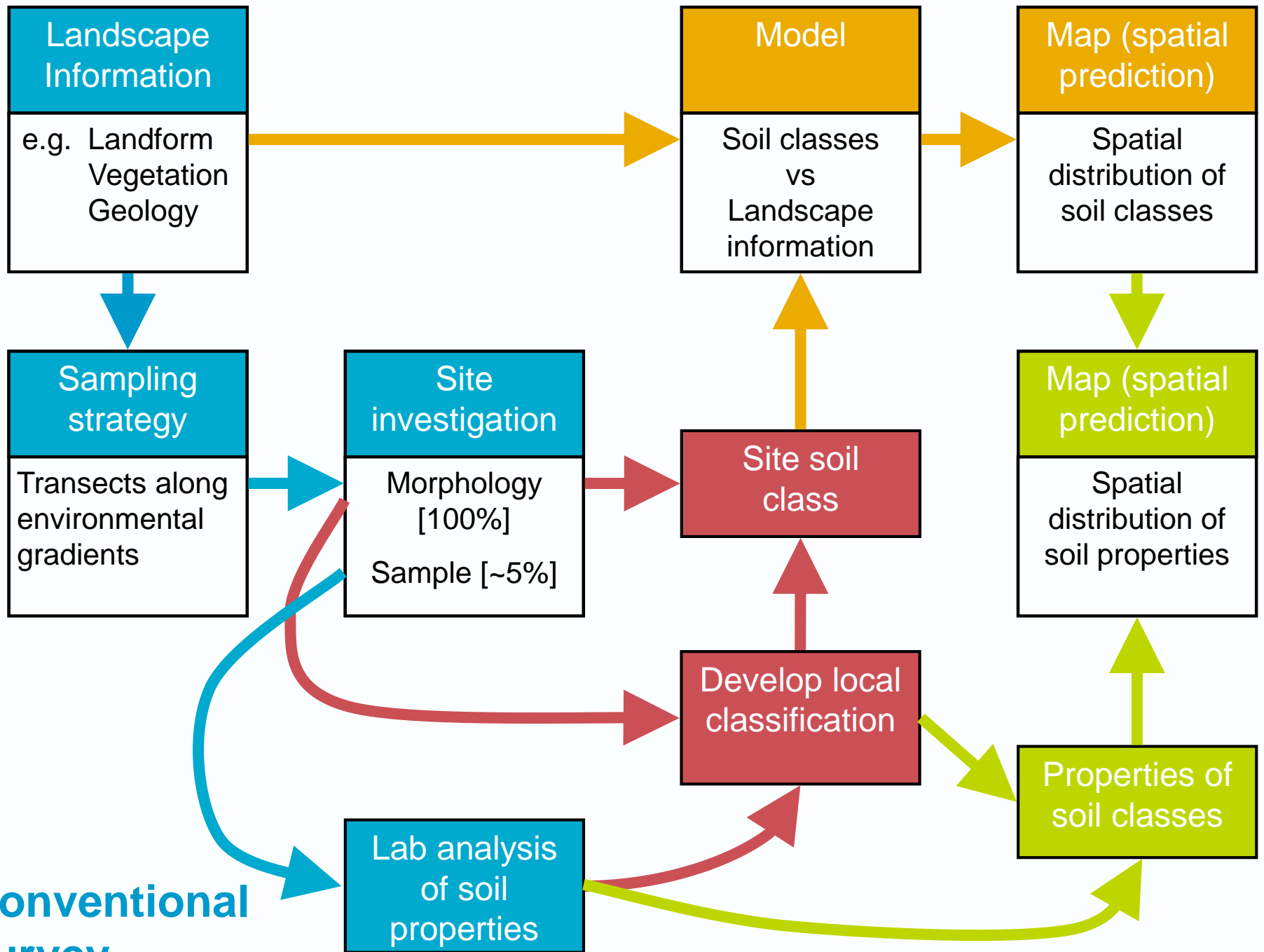
Provides:

- Quantitative data
- Unlike morphology, measures soil properties of interest to farmers, hydrologists, land use planners etc.

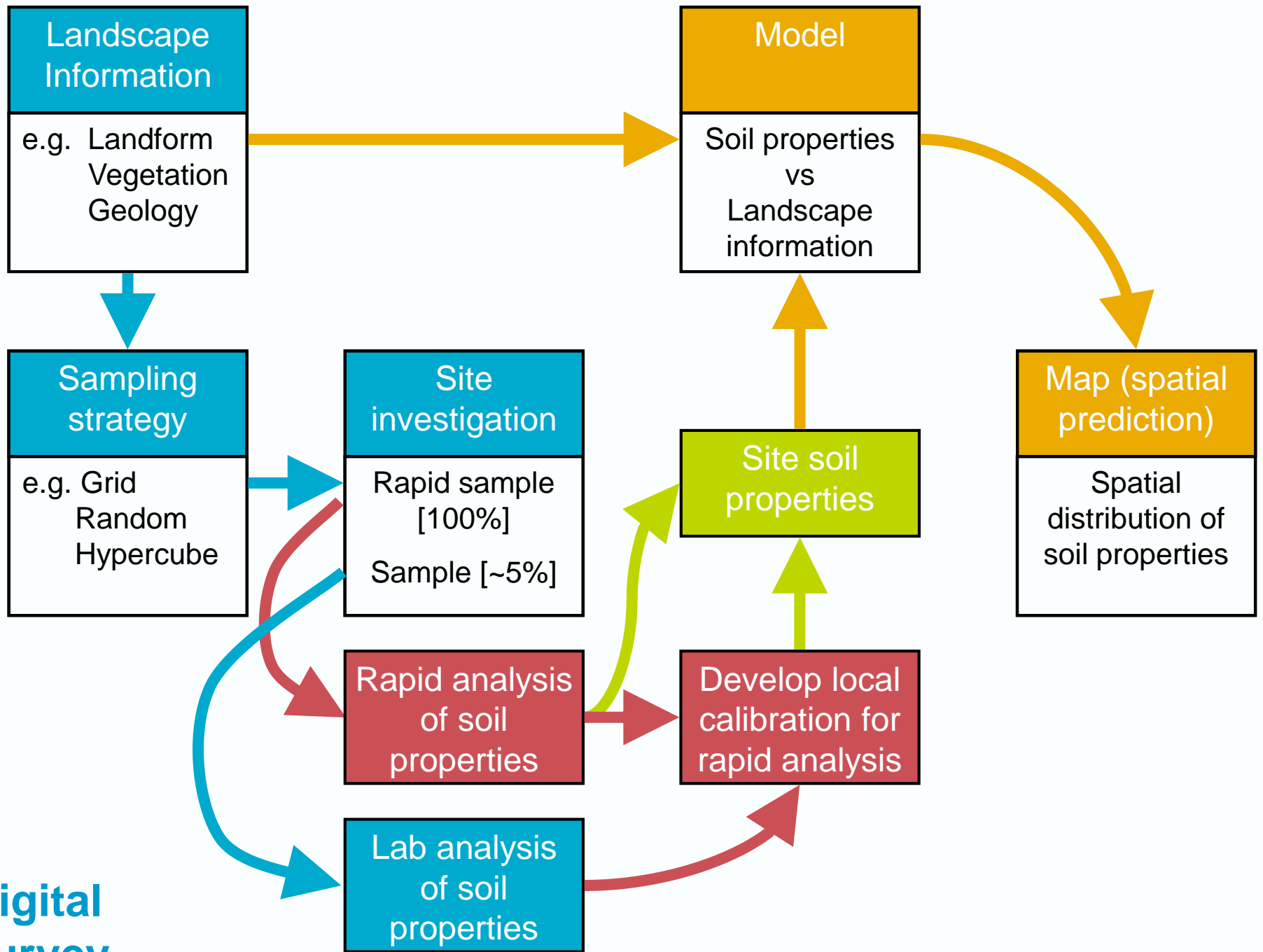
Not as precise as laboratory measurements

Measurement strategy for soil survey:

- Many, less precise measurements  
are better than  
few precise measurements
- **Because spatial variability of soil properties is large**
  - **Even over short distances**



**Conventional survey**



# Digital Land Resource Mapping (DLRM)

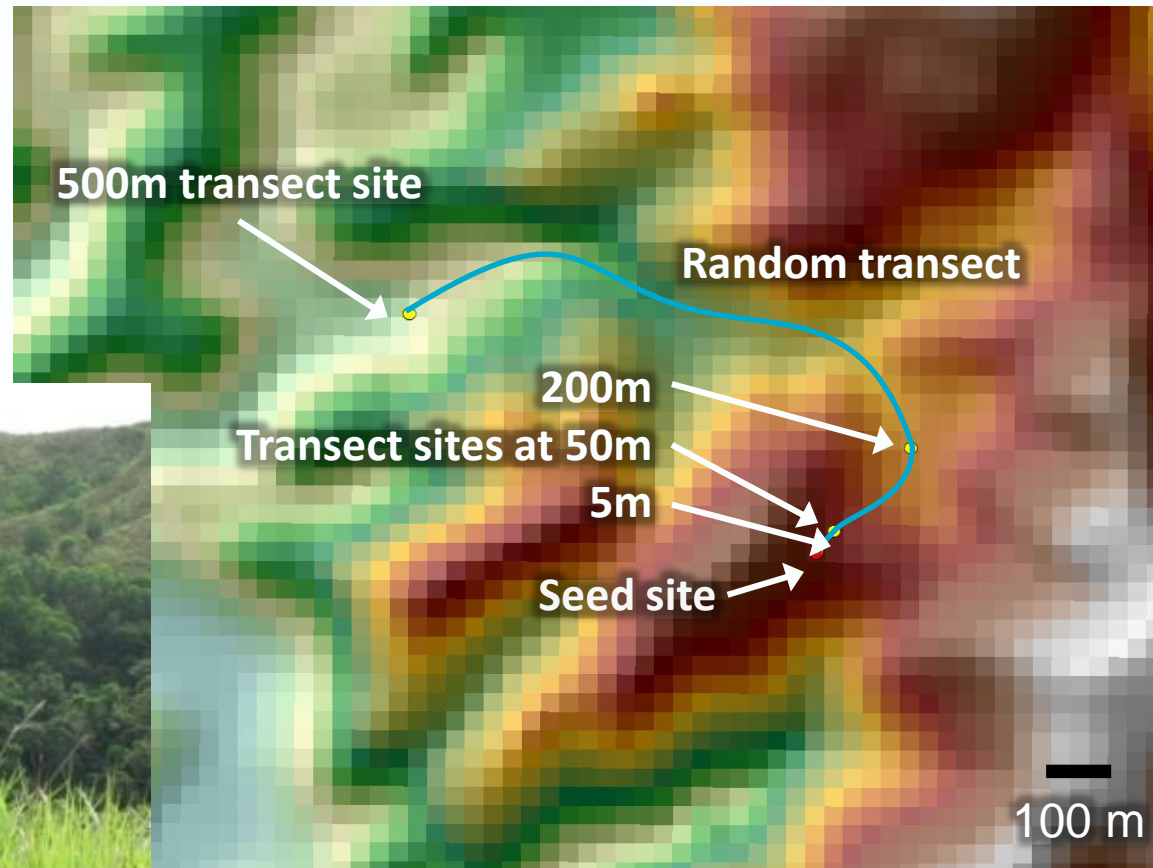
including Digital Soil Mapping (DSM)

Based on 4 pillars:

1. Statistically-based sampling strategy
2. Simplified site and soil profile methods
3. Rapid soil analysis
4. Mapping of soil and land properties using statistical models

# 1. Statistically-based sampling strategy

Using random transect samples generated on a 30 m digital elevation model (DEM)





# 2. Simplified site and soil profile methods

Simplified field procedures that can be carried out by non-experts with an emphasis on soil sampling all sites





# 3. Rapid soil analysis

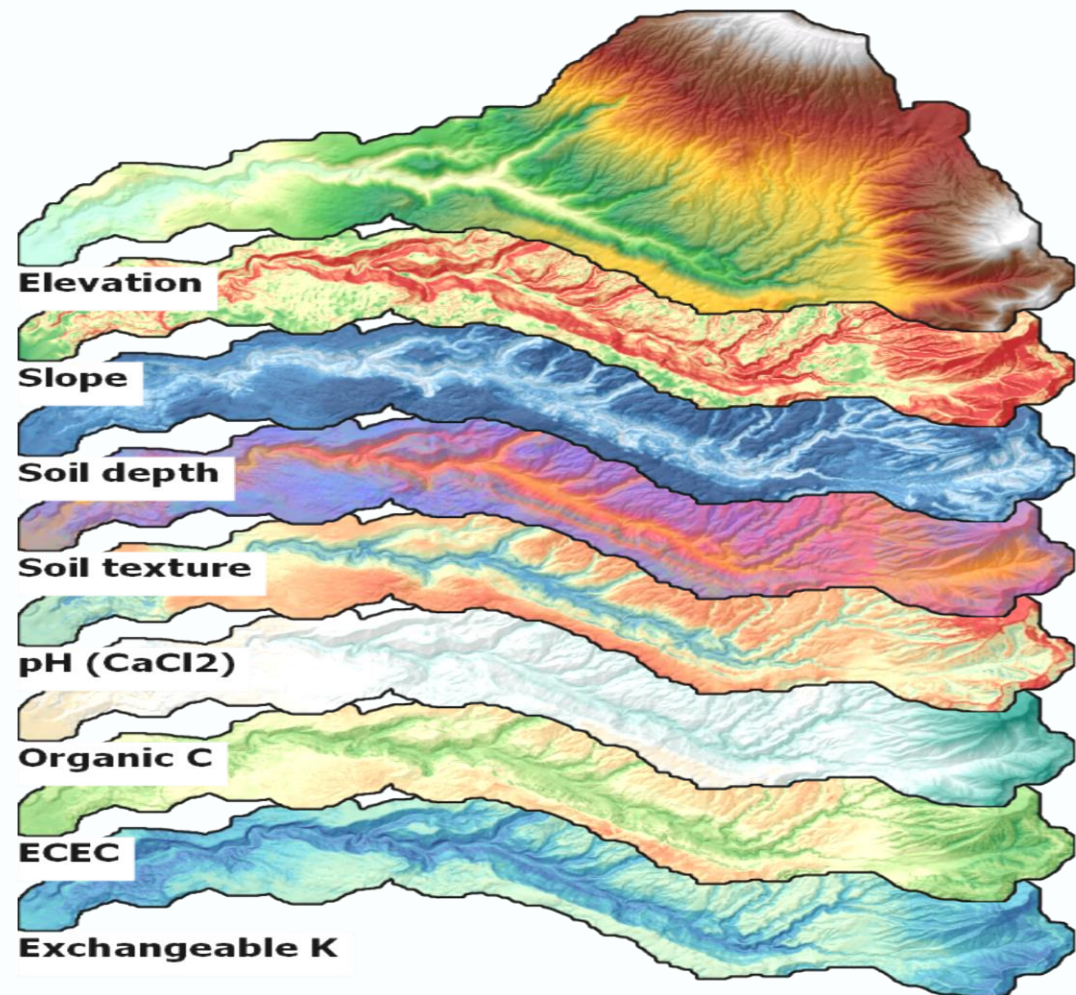
- Uses mid-infrared spectroscopy (MIR)
- Possible to estimate a range of soil properties at all sample sites
  - rather than at a small subset when only conventional laboratory methods are used.



# 4. Mapping of soil and land properties using statistical models

Numerical methods are used

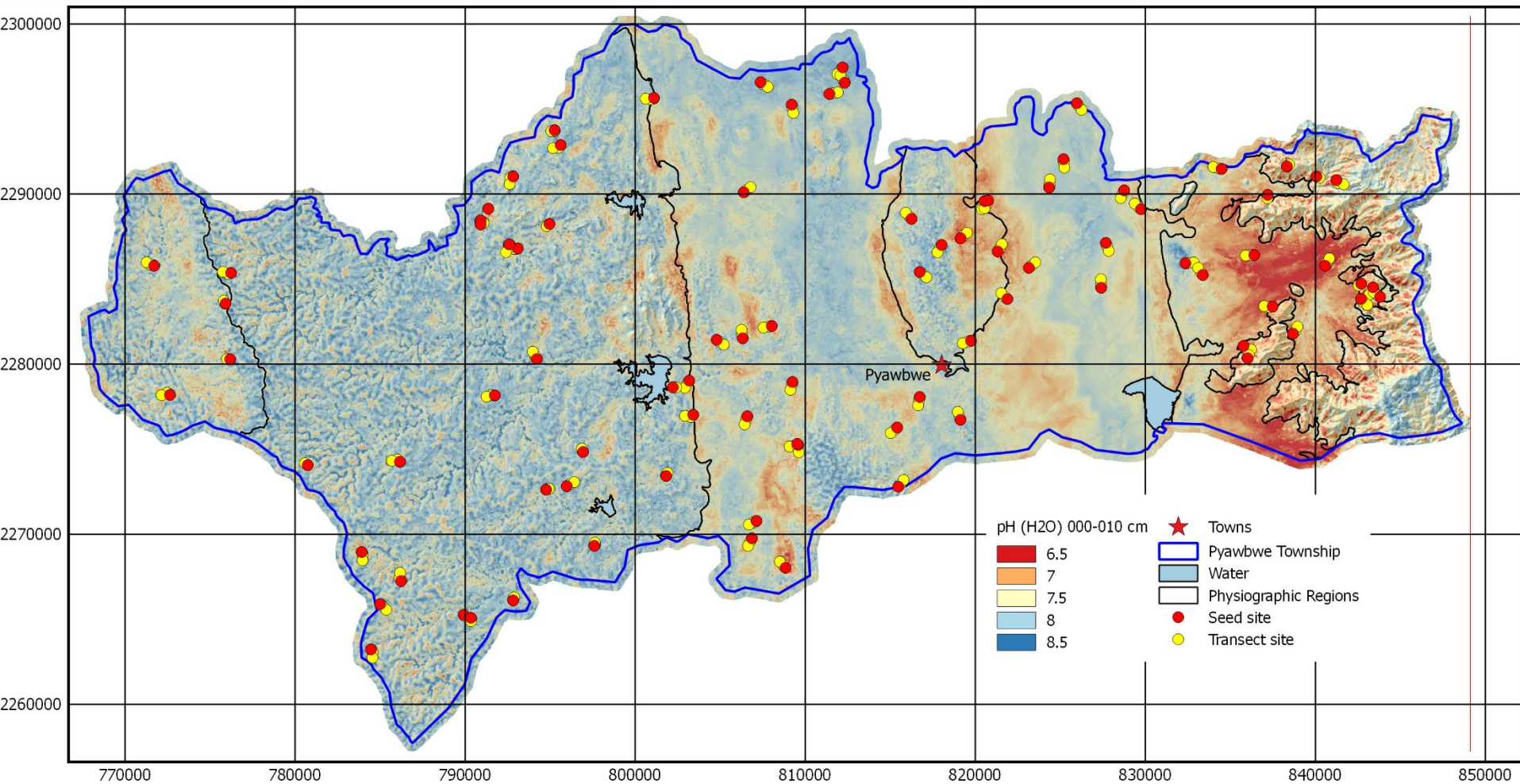
- To predict land and soil properties over the survey area using
  - Measurements at sample sites
  - Terrain attributes (e.g. slope) derived from a 20 m digital elevation model (DEM)
- Produces, not one, but many digital maps such as these



*Cabulig watershed,  
southern Philippines*

# Digital soil map of Pyawbwe Township

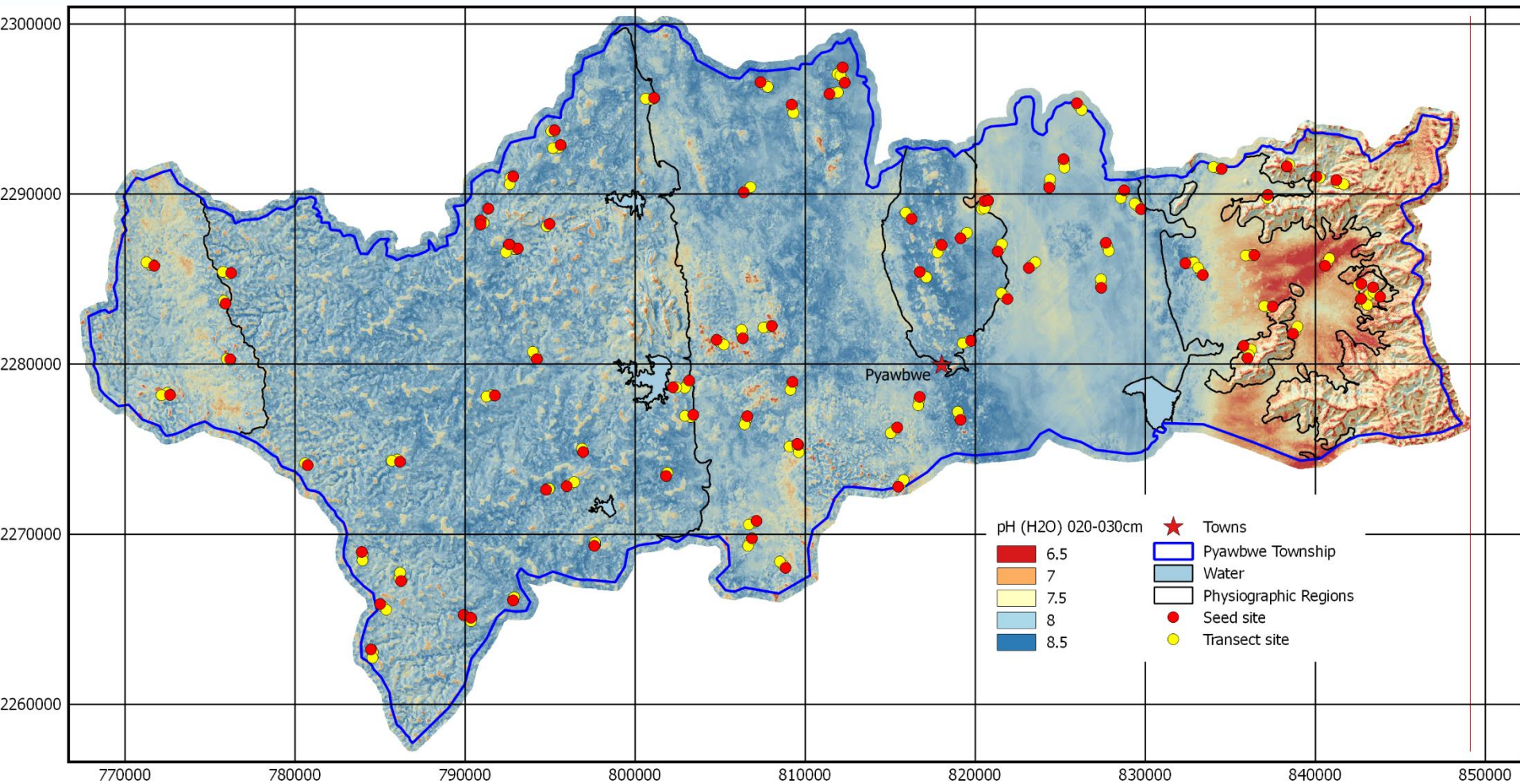
pH (1:5 H<sub>2</sub>O) at 0 - 10 cm depth





# Digital soil map of Pyawbwe Township

pH (1:5 H<sub>2</sub>O) at 20 - 30 cm depth



# Conclusion

## Rapid soil analysis by MIR spectroscopy

- Allows **direct** mapping of soil properties of interest to land managers
- Instead of mapping soil type based on observations of soil morphology and then **inferring** soil properties from a measurements on a few examples of each soil type



# Rapid Soil Analysis by Mid-Infrared Spectroscopy Training Course

## 04. SAFETY

Anthony Ringrose-Voase  
May 2018

Agriculture and Food  
[www.csiro.au](http://www.csiro.au)

# Health and safety

Highest priority objective for any project:

- That all staff return home
- Uninjured
- In good health

Outline of presentation

- CSIRO framework for assessing health and safety risks
- Controlling risk
- Risk assessment for field laboratory

# Framework for assessing safety

## 1. Identify a hazard

- For example, repetitive strain injury due to soil grinding



# Framework for assessing safety

## 2. Assess the **consequence** of the hazard

- Catastrophic
  - Fatality
  - Serious, permanent disability
- Major
  - Extended absence from work (>1 week)
  - Moderate to severe health effects
- Moderate
  - Temporary absence from work (<1 week)
  - Requires one or more visits to doctor
- Minor
  - Requires first aid
- Insignificant
  - No injury

# Framework for assessing safety

## 3. Assess **likelihood** of hazard

- Almost certain
  - Daily to several times per week
- Likely
  - Once per month to several times per year
- Possible
  - Once per year to several times over 2 years
- Unlikely
  - Once every 2 to 5 years
- Rare
  - Once every 5 years or less

# Framework for assessing safety

## 4. Assess inherent risk

- Risk that exists in the absence of control measures
- Assessed using consequence and likelihood

Likelihood	Consequence				
	Insignificant	Minor	Moderate	Major	Catastrophic
Almost certain	Mod	Mod	High	High	High
Likely	Mod	Mod	Mod	High	High
Possible	Low	Mod	Mod	High	High
Unlikely	Low	Low	Mod	Mod	High
Rare	Low	Low	Mod	Mod	Mod

# Framework for assessing safety

## Inherent risk levels

### High risk

- Immediate action required

### Moderate risk

- Management responsibility must be specified

### Low risk

- Manage by routine procedures

# Framework for assessing safety

## 5. Decide on management controls

- Very good
  - Controls are best practice
  - Involve explicit standards
  - Followed all of the time
  - High emphasis on elimination, substitution or engineering controls.
- Reasonable
  - Controls are in place
  - Not followed all of the time
  - May not include best practice
  - High emphasis on administration and protective equipment.
- Poor
  - Few or no controls in place.
  - No standards have been identified.
  - Controls do not address Hierarchy of Control principles.

# Framework for assessing safety

## 6. Assess impact of management controls

- Determines residual risk
- Risk that exists after application of control measures

Likelihood	Consequence				
	Insignificant	Minor	Moderate	Major	Catastrophic
Almost certain	Mod	Mod	High	High	High
Likely	Mod	Mod	Mod	High	High
Possible	Low	Mod	Mod	High	High
Unlikely	Low	Low	Mod	Mod	High
Rare	Low	Low	Mod	Mod	Mod

# Framework for assessing safety

## Residual risk levels

- **High** Attend to immediately
- **Moderate** Attend to in short-term
- **Low** Attend to in longer term

# Hierarchy of control principles

## Elimination

- Eliminate the hazard from the workplace altogether

## Substitution/modification

- Substitute the hazard with something with lesser risk which will still perform the same task in a satisfactory manner

## Isolation

- Isolate the problem from staff
- often by use of separate purpose built rooms, barricades, sound barriers, etc.

## Engineering controls

- Re-design equipment, work processes or tools to reduce or eliminate the risk

Most  
preferred  
controls



# Hierarchy of control principles

## Administrative controls

- Training
- Written work procedures
- Adequate supervision
- Signage
- Maintenance of plant and equipment
- Limitation of exposure time

## Personal protective equipment (PPE)

- Provide adequate personal protective equipment.
- Masks, gloves, boots, overalls etc.

### Note:

- This is the least preferable method of dealing with a hazard
- It should only be adopted when all other methods are unsuitable.



Least preferred controls

# Exercise: Risk assessment for MIR laboratory

## Exercise:

1. Identify a hazard you think we might encounter in the field laboratory  
Then work through the risk assessment framework for that hazard
2. Consequences
3. Likelihood
4. Inherent risk
5. Management controls
6. Residual risk

# Hazards in MIR laboratory

## Repetitive strain injury

- Long term pain due to repetitive actions during grinding
- **Consequence:** Major
- **Likelihood:** Likely
- **Inherent risk:** High
- **Controls:**
  - Staff should be aware of risk and of need to act early
  - Provide ergonomic workstations
  - Regular rest-breaks – rotate activity with other staff
- **Consequence:** Moderate
- **Likelihood:** Unlikely
- **Residual risk:** Moderate

# Hazards in MIR laboratory

## Electrocution

- Use of electric equipment – oven, MIR spectrometer
- Serious injury, death
- **Consequence:** Catastrophic
- **Likelihood:** Unlikely
- **Inherent risk:** High
- Controls:
  - Use earth
  - Check electric leads for wear and tear
  - Maintain equipment
  - Use approved plugs and sockets
  - Use licensed electrician to change plugs etc.
- **Consequence:** Catastrophic
- **Likelihood:** Rare
- **Residual risk:** Moderate



# Rapid Soil Analysis by Mid-Infrared Spectroscopy Training Course

## 05. MIR CALIBRATION STRATEGY

Anthony Ringrose-Voase

May 2018

Agriculture and Food  
[www.csiro.au](http://www.csiro.au)



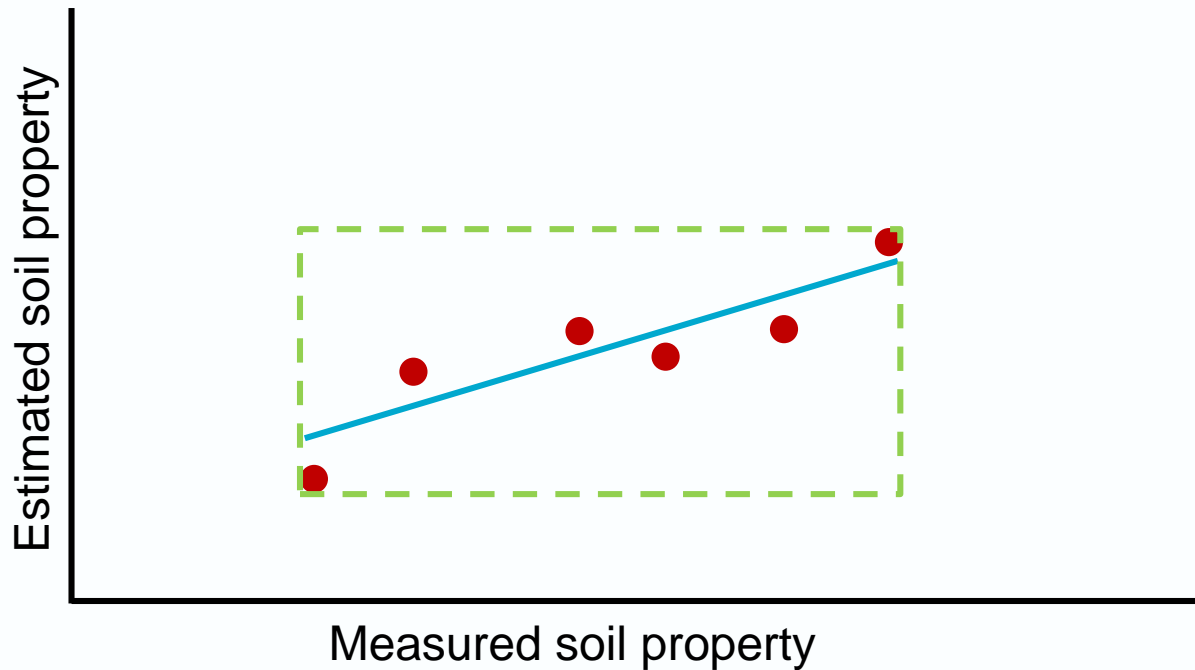
Australian Government  
Australian Centre for  
International Agricultural Research



# Choice of samples for calibration

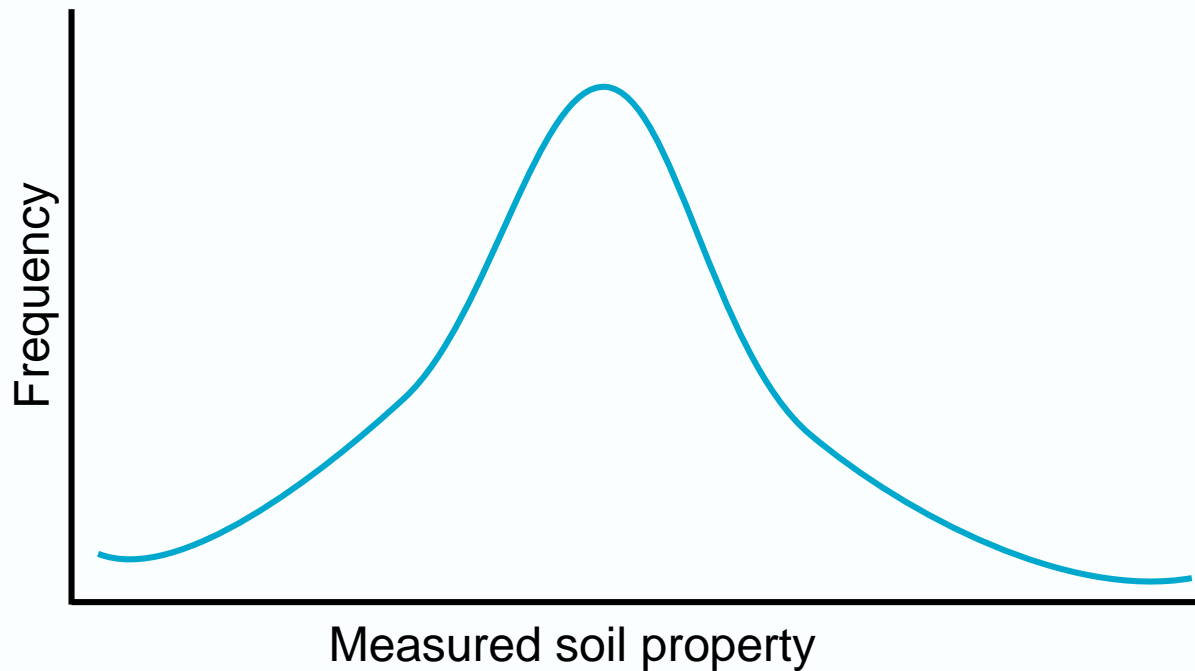
Prediction of a soil property using a calibration is only valid for soil property values within the range of the calibration sample set

- Properties of calibration samples should cover full range encountered in survey area



# Choice of samples for calibration

Calibrations are most reliable when statistical distribution of soil properties in the calibration sample set reflects the distribution in the survey area



# Calibration strategy for a survey area

- At the outset we have no knowledge of the range or distribution of soil properties in survey area
- Aim to represent the range of soil materials in survey area
- Two stage process
  - Select sampling locations likely to include a wide range of soil materials
  - After acquiring MIR spectra, select samples that cover the range of MIR spectral characteristics



# Calibration strategy for a survey area

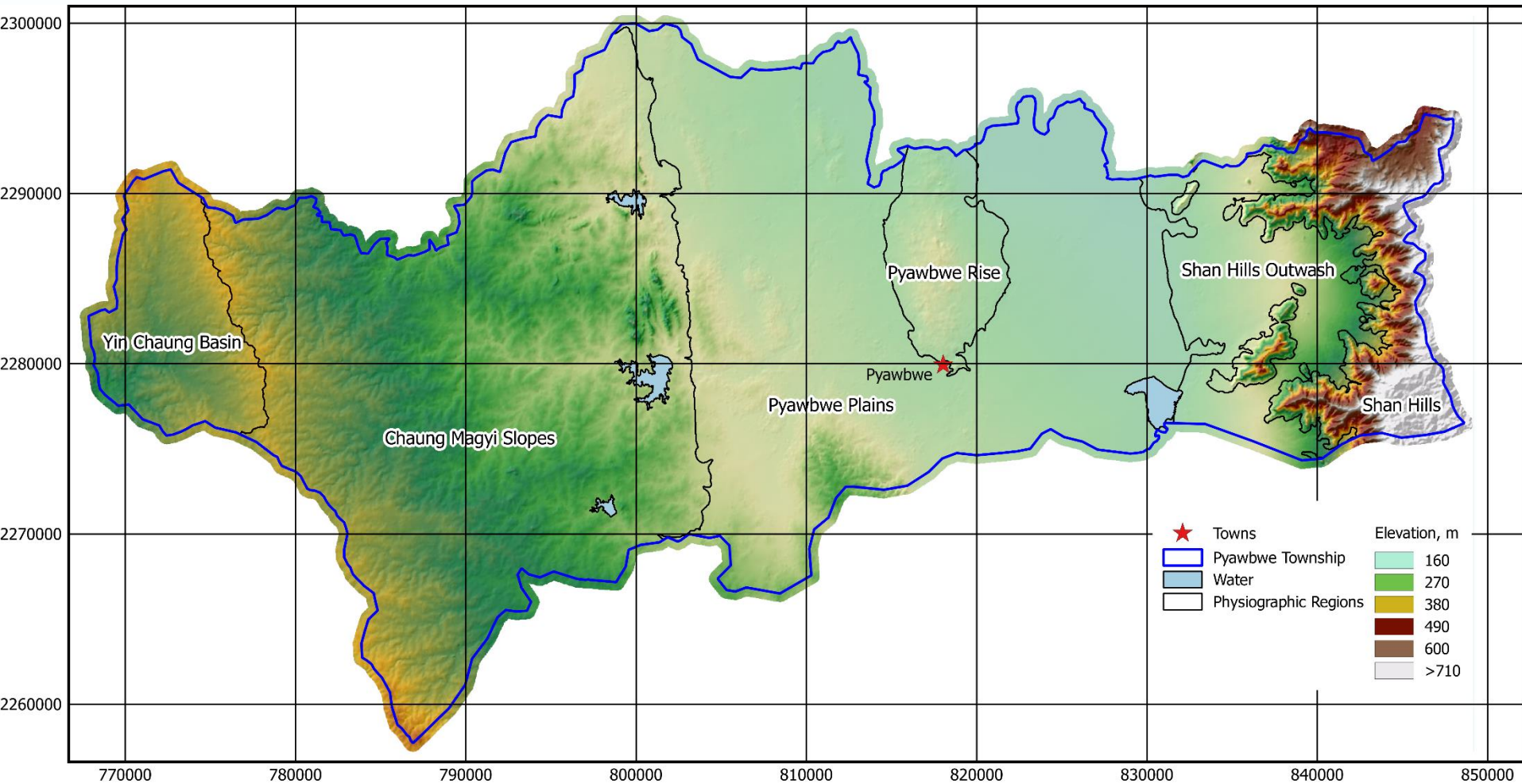
## Stage 1

Select sampling locations likely to include a wide range of soil materials

- Stratify landscape at two levels
  - Physiographic regions
  - Landform units

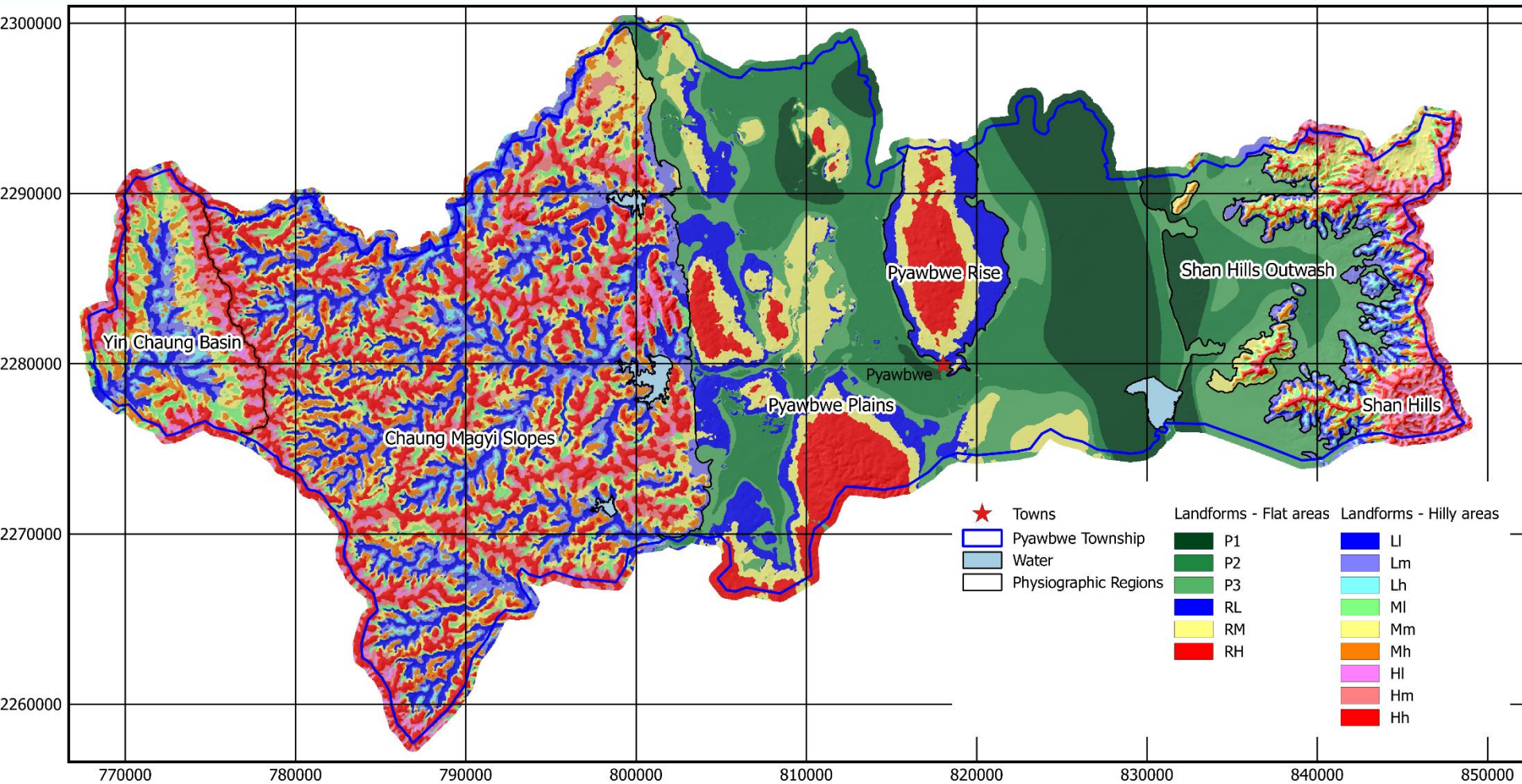
# Calibration strategy for a survey area

## Stage 1 – Stratification into physiographic regions



# Calibration strategy for a survey area

## Stage 1 – Stratification of each physiographic region into landform units



# Calibration strategy for a survey area

## Stage 1

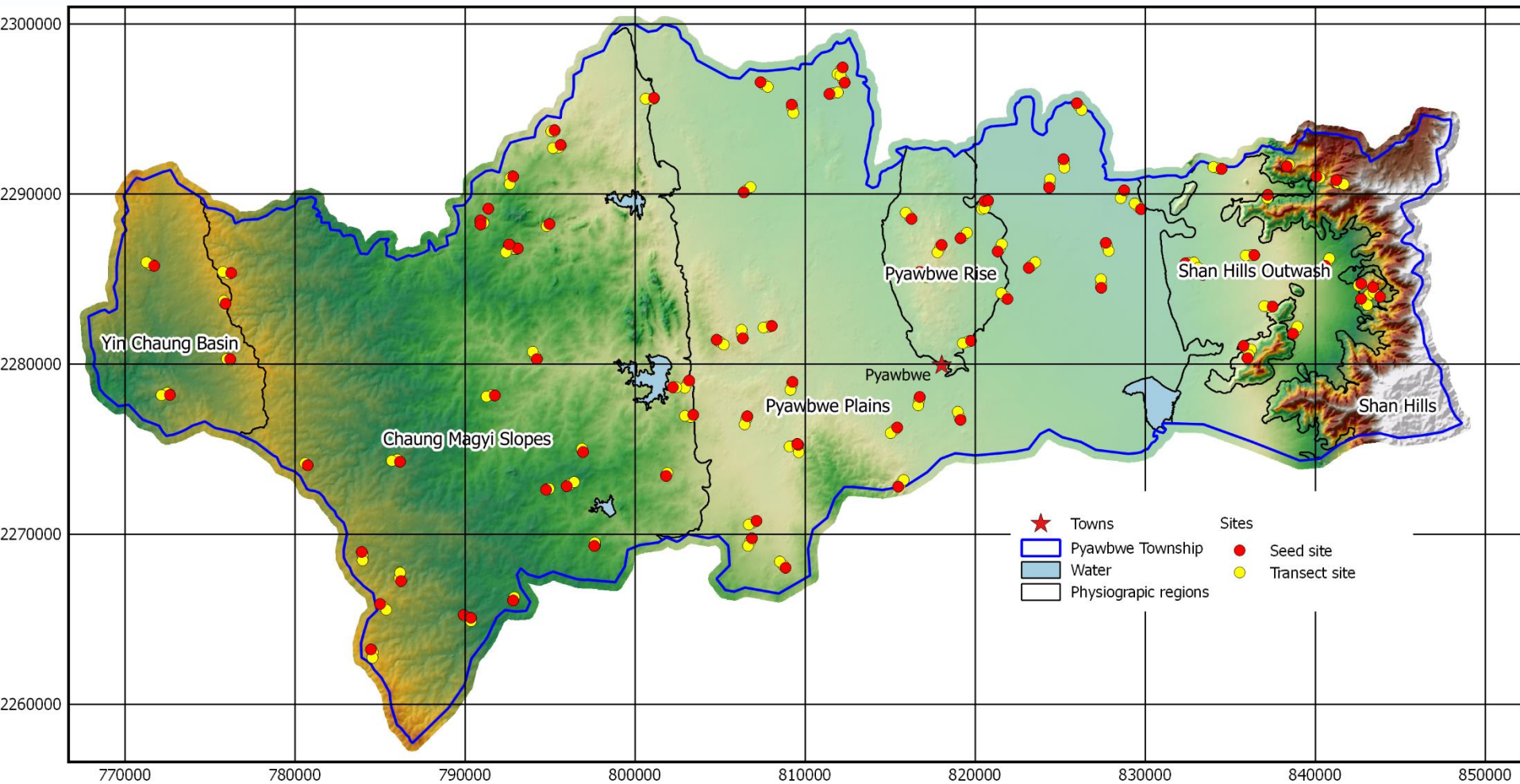
Select sampling locations likely to include a wide range of soil materials

- Stratify landscape at two levels
  - Physiographic regions
  - Landform units
- Randomly locate 'seed sites' in each physiographic-landform unit
  - Number of sites proportional to unit area
- Also locate 4 'transect sites' along random transects away from seed site



# Calibration strategy for a survey area

## Stage 1 – Stratified-random sampling strategy



# Calibration strategy for a survey area

## Stage 1

Select sampling locations likely to include a wide range of soil materials

- Stratify landscape at two levels
  - Physiographic regions
  - Landform units
- Randomly locate 'seed sites' in each physiographic-landform unit
  - Number of sites proportional to unit area
- Also locate 4 'transect sites' along random transects away from seed site
- Specimen collection
  - Seed sites: MIR specimen + bag specimen (Pyawbwe – 498 specimens)
  - Transect sites: MIR specimen only (Pyawbwe – 3561 specimens)

(Pyawbwe – 4059 total)

# Calibration strategy for a survey area

## Stage 2

From seed site specimens select specimens that cover the range of MIR spectral characteristics

- Scan all MIR specimens from seed sites
  - (Pyawbwe – 498 specimens)
- Analyse MIR spectra
  - Using principal component analysis
- Select a sub set of ~100 specimens that cover range of principal components
  - Using Kennard Stone sampling method

# Calibration strategy for a survey area

- Conventional analysis of bag specimens for selected MIR spectra
  - 104 specimens (out of all seed site specimens – 498 in Pyawbwe)
- Develop calibrations for each soil property of interest
- Scan MIR specimens from transect sites
  - (3561 specimens in Pyawbwe)
- Estimate soil properties for unanalysed specimens using calibrations
  - Pyawbwe: 3561 transect specimens + 394 seed specimens = 3955 specimens
- Outliers with unreliable estimates are automatically identified
  - Can be removed if desired
  - Or sent for conventional analysis to update calibration



# Types of calibration strategies

- Survey area calibrations
  - As above (e.g. for township, catchments)
- Regional calibrations
  - E.g. Central Dry Zone
  - Start with calibration for a survey area
  - Gradually add sites for other areas in the region
  - Incrementally update calibrations
- National calibration
  - Use: Existing database of soil test values within a country  
Archive of soil specimens (from surveys, agronomy trials etc.)
- Global calibration

# Types of calibration strategies

General rule:

- Local calibrations will generally give more accurate estimates than regional, national or global calibrations
  - But are limited to the calibration area
- Spectra should be archived together with soil specimens
  - Allows calibrations to be developed for new soil properties in the future
- Most effective if different institutions share spectra and results to build up databases of soil properties and MIR spectra



# Rapid Soil Analysis by Mid-Infrared Spectroscopy

## Training Course

### 06. MIR CALIBRATION

Anthony Ringrose-Voase

May 2018

Agriculture and Food

[www.csiro.au](http://www.csiro.au)



Australian Government

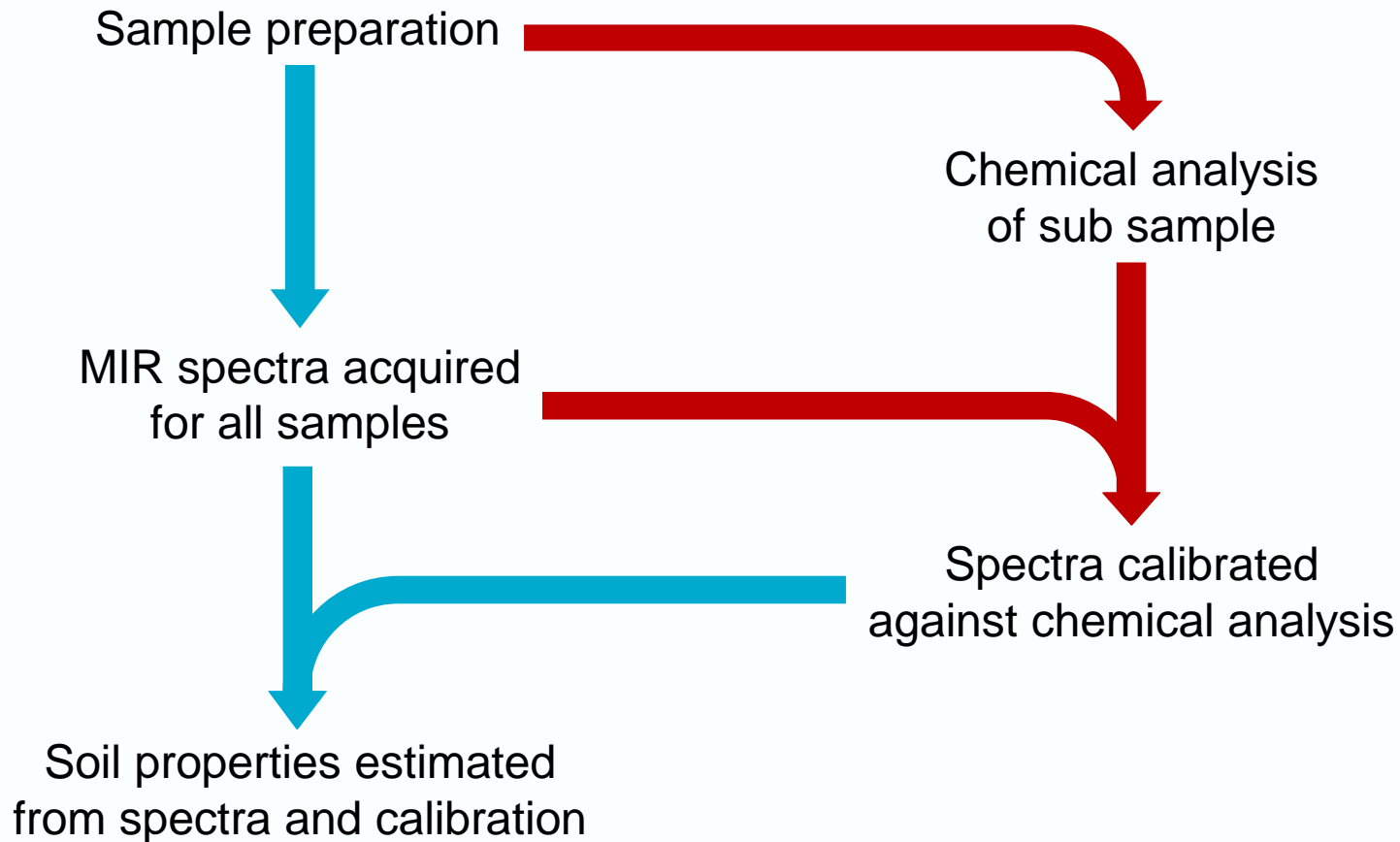
Australian Centre for  
International Agricultural Research



# Purpose of MIR calibration

- To correlate measured MIR spectra with soil properties measured by conventional analysis
- To use this correlation to estimate these properties for soil samples for which there is no conventional analysis
- The whole spectrum is used in correlation
- The correlation is done by partial least square regression

# MIR Process to measure soil properties

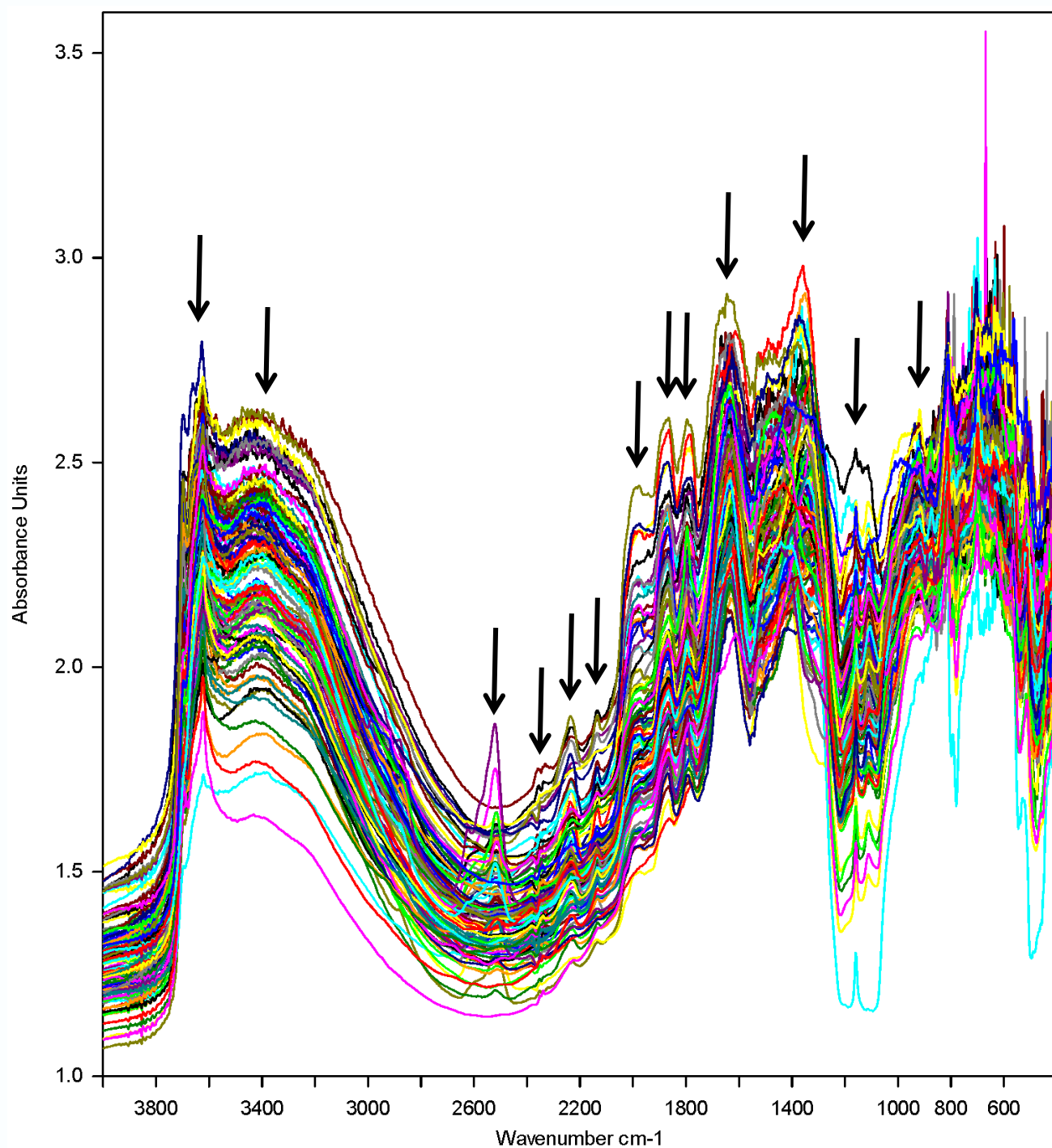


# MIR spectra of 104 samples

Pyawbwe Township,  
Myanmar

Many parts of the  
spectra follow a  
similar pattern

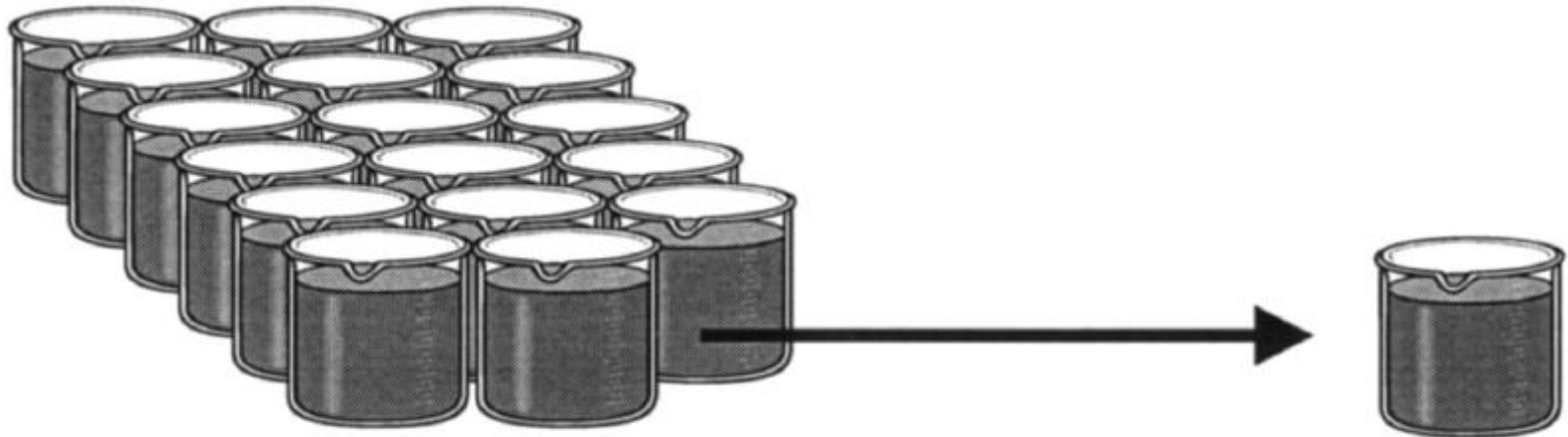
- Therefore much  
is redundant  
information
- The whole  
spectra can be  
described by a  
few principal  
components



# Cross-validation

## Checking the quality of a calibration

- Calibration is run many times
  - Each time one sample is removed and used for validation
- Good method when short of samples: minimum of  $\sim 60$  required for a calibration



Calibration Set:  
Developing a Method

Test Sample:  
Validating the Method

# Residual prediction deviation (RPD)

## Assessment of calibration

- RPD = Standard deviation/Root mean square error of prediction
- Related to  $R^2$
- Indicates quality of prediction:

< 1.5	Poor
1.5-2.0	Indicator
2.0-3.0	Good
> 3.0	Analytical quality



# MIR prediction for soil samples

## Cabulig watershed, southern Philippines

Soil attribute	RPD	Prediction quality
Organic carbon	3.1	Analytical quality
pH (1:5 H <sub>2</sub> O)	2.8	Good quality
Exchangeable Mg	2.7	Good quality
Exchangeable acid	2.3	Good quality
Sum exchangeable cation	2.3	Good quality
Clay content	2.2	Good quality
Exchangeable Ca	2.2	Good quality
ECEC	2.0	Good quality
Sand content	1.8	Indicator
Exchangeable K	1.5	Indicator
Silt	1.4	Poor
Olsen P	1.3	Poor

# MIR prediction for soil samples

## Inabanga watershed, southern Philippines

Soil attribute	RPD	Prediction quality	(Cabulig)
Organic carbon	2.1	Good quality	(Analytical quality)
pH (1:5 H <sub>2</sub> O)	2.0	Good quality	(Good quality)
Exchangeable Mg	2.1	Good quality	(Good quality)
Exchangeable acid	1.7	Indicator	(Good quality)
Sum exchangeable cation	3.8	Analytical quality	(Good quality)
Clay content	1.8	Indicator	(Good quality)
Exchangeable Ca	3.4	Analytical quality	(Good quality)
ECEC	3.4	Analytical quality	(Good quality)
Sand content	1.7	Indicator	(Indicator)
Exchangeable K	1.6	Indicator	(Indicator)
Silt	1.4	Poor	(Poor)
Olsen P	1.1	Poor	(Poor)

# Calibration process

- Make a table of spectra of calibration samples
- Make a table of laboratory results
- Check match of calibration and lab analysis
- Calculate calibration
- Validate
- Optimize



# Rapid Soil Analysis by Mid-Infrared Spectroscopy Training Course

## 07. MIR CALIBRATION INSTRUCTIONS

Anthony Ringrose-Voase

May 2018

Agriculture and Food  
[www.csiro.au](http://www.csiro.au)



Australian Government  
Australian Centre for  
International Agricultural Research



# MIR Calibration with OPUS Quant 2 module

## Start Quant 2

The screenshot displays the OPUS software interface. The main window is titled "OPUS - Operator: Administrator (Administrator) - [Display - alpha.ows]". The menu bar includes File, Edit, View, Window, Measure, Manipulate, Evaluate, Display, Print, Macro, Validation, Setup, and Help. The toolbar contains various icons for file operations, measurement, and analysis. A red circle highlights an icon in the toolbar that resembles a balance scale, with the coordinates  $(2539.163, 0.7075)$  displayed next to it. The main display area is a graph with a vertical axis ranging from 0.000 to 1.500 and a horizontal axis ranging from 4000 to 600. The graph is currently blank. The bottom status bar shows "Info" and "No Active Task". The Windows taskbar at the bottom includes icons for the Start menu, File Explorer, Mail, OneDrive, Google Chrome, and PowerPoint, along with system tray icons for volume, network, and battery, and the date/time "7:04 PM 7/05/2018".

# First create a calibration template

- Template will contain:
  - List of soil properties for which there are calibration data
  - List of calibration spectra
    - How many replicate spectra there are for each sample
  - Calibration data
  - Other calibration settings
- Setting up a template correctly makes it easier to perform the calibrations in a standard way



Load Method **Components** Spectra Parameters Validate Graph Report Store Method Optimize Settings

Add Component

Name

Unit

Formatting in the Quant 2 analysis report

Default settings (5 significant digits)

Digits after the decimal point



- Load Method
- Components
- Spectra
- Parameters
- Validate
- Graph
- Report
- Store Method
- Optimize
- Settings

1. Add Component

Name	Unit
2. pH_H2O	3.

- pH\_H2O

Formatting in the Quant 2 analysis report

- Default settings (5 significant digits)
- 4.  Digits after the decimal point

2





Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

Add Component

Name

Unit

pH\_H2O

pH\_H2O

EC [dS/m]

OC [%]

P\_Olsen [ppm]

PBI

Exch\_Ca [cmol+/kg]

Exch\_Mg [cmol+/kg]

Exch\_K [cmol+/kg]

Exch\_Na [cmol+/kg]

Exch\_Acid [cmol+/kg]

Exch\_Sum [cmol+/kg]

ECEC [cmol+/kg]

BaseSat [%]

ESP [%]

Ca\_Mg

ECEC\_Clay

Sand [%]

Silt [%]

Clay [%]

In\_Sand\_Clay

Formatting in the Quant 2 analysis report

Default settings (5 significant digits)

Digits after the decimal point

2



Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

Add Spectra

Change Path

Window

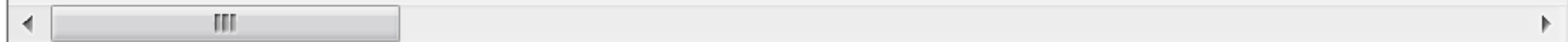
Set Sample Numbers

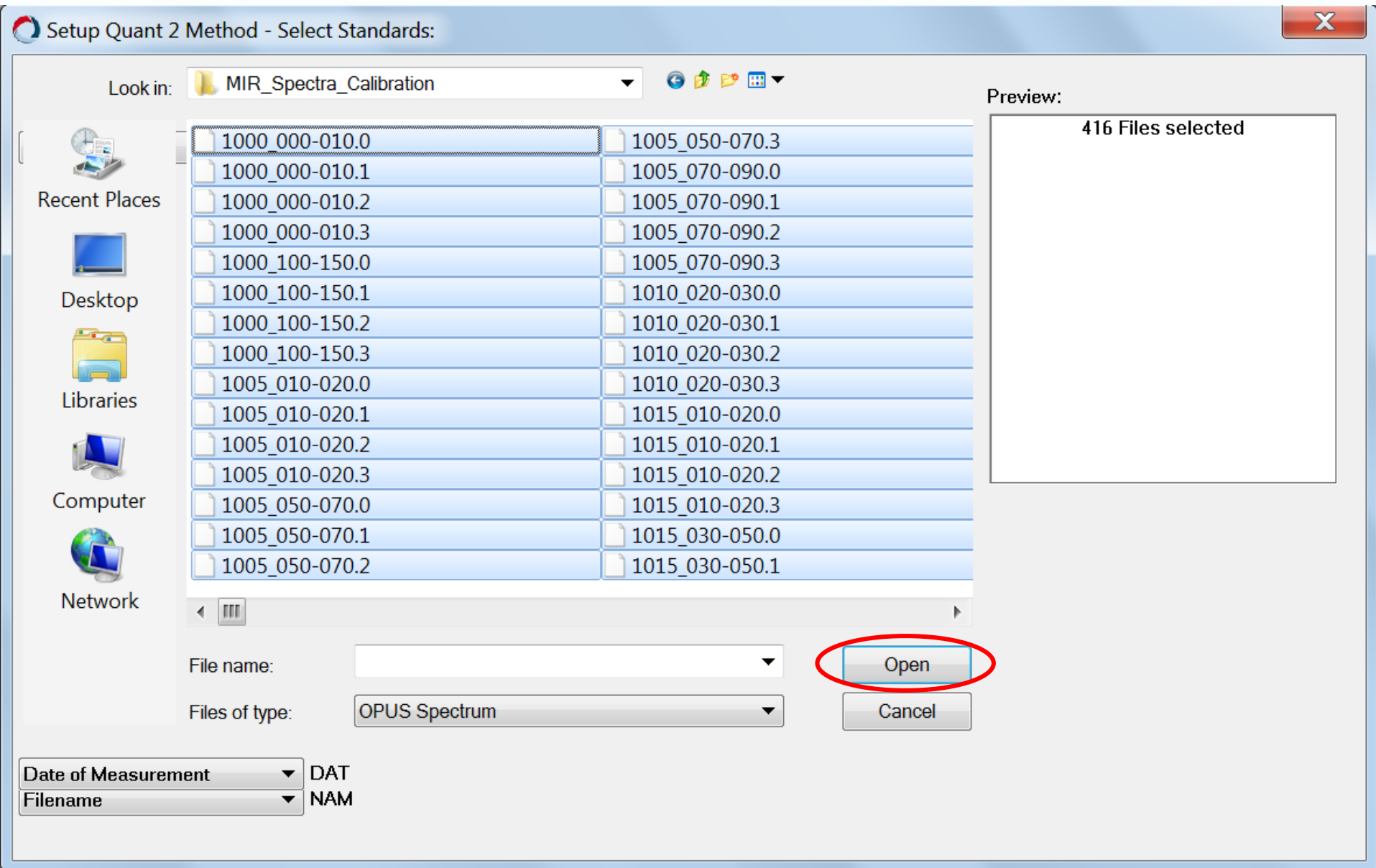
Set Data Set

Print

	Data Set	Sample	Path	File Name	pH_H2O	EC	OC	P_Olsen	PBI	Excl
--	----------	--------	------	-----------	--------	----	----	---------	-----	------

--	--	--	--	--	--	--	--	--	--	--





■ Hint: Click first spectra then <Ctrl A> to select all spectra in folder



Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

Add Spectra

Change Path

Copy Spectra

Window

Set Sample Numbers

Set Data Set

Comp. Correlations

Print

	Data Set	Sample	Path	File Name	pH_H2O	EC	OC	P_Olsen	PBI	▲
1	Calibration	1	C:\Users\rin0	1000_000-010.0						☰
2	Calibration	1	C:\Users\rin0	1000_000-010.1						
3	Calibration	1	C:\Users\rin0	1000_000-010.2						
4	Calibration	1	C:\Users\rin0	1000_000-010.3						
5	Calibration	2	C:\Users\rin0	1000_100-150.0						
6	Calibration	2	C:\Users\rin0	1000_100-150.1						
7	Calibration	2	C:\Users\rin0	1000_100-150.2						
8	Calibration	2	C:\Users\rin0	1000_100-150.3						
9	Calibration	3	C:\Users\rin0	1005_010-020.0						
10	Calibration	3	C:\Users\rin0	1005_010-020.1						
11	Calibration	3	C:\Users\rin0	1005_010-020.2						
12	Calibration	3	C:\Users\rin0	1005_010-020.3						
13	Calibration	4	C:\Users\rin0	1005_050-070.0						
14	Calibration	4	C:\Users\rin0	1005_050-070.1						
15	Calibration	4	C:\Users\rin0	1005_050-070.2						
16	Calibration	4	C:\Users\rin0	1005_050-070.3						
17	Calibration	5	C:\Users\rin0	1005_070-090.0						
18	Calibration	5	C:\Users\rin0	1005_070-090.1						
19	Calibration	5	C:\Users\rin0	1005_070-090.2						
20	Calibration	5	C:\Users\rin0	1005_070-090.3						
21	Calibration	6	C:\Users\rin0	1010_020-030.0						▼

### Set Sample Numbers

Number of spectra per sample:

1

1.  Set sample numbers according to file names

2.

3.

Spectra

Window

Correlations

Print

	D	OC	P_Olsen	PBI
1	Cali			
2	Cali			
3	Cali			
4	Cali			
5	Cali			
6	Cali			
7	Cali			
8	Cali			
9	Cali			
10	Cali			
11	Calibration			
12	Calibration	3	C:\Users\rin0	1005_010-020.3
13	Calibration	4	C:\Users\rin0	1005_050-070.0
14	Calibration	4	C:\Users\rin0	1005_050-070.1
15	Calibration	4	C:\Users\rin0	1005_050-070.2
16	Calibration	4	C:\Users\rin0	1005_050-070.3
17	Calibration	5	C:\Users\rin0	1005_070-090.0
18	Calibration	5	C:\Users\rin0	1005_070-090.1
19	Calibration	5	C:\Users\rin0	1005_070-090.2
20	Calibration	5	C:\Users\rin0	1005_070-090.3
21	Calibration	6	C:\Users\rin0	1010_020-030.0

- In Excel ensure that any missing values are BLANK
- Copy calibration data in Excel
- Paste into OPUS
- CHECK that the data is lined up against the correct sample names
- Check again
- And again
  - It is really important that OPUS matches the correct data with each spectrum



Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

Add Spectra

Change Path

Copy Spectra

Window

Set Sample Numbers

Set Data Set

Comp. Correlations

Print

	Data Set	Sample	Path	File Name	pH_H2O	EC	OC	P_Olsen	PBI	▲
1	Calibration	1	C:\Users\rin0	1000_000-010.0	7.85	0.224	1.85	52.28	93	☰
2	Calibration	1	C:\Users\rin0	1000_000-010.1	7.85	0.224	1.85	52.28	93	
3	Calibration	1	C:\Users\rin0	1000_000-010.2	7.85	0.224	1.85	52.28	93	
4	Calibration	1	C:\Users\rin0	1000_000-010.3	7.85	0.224	1.85	52.28	93	
5	Calibration	2	C:\Users\rin0	1000_100-150.0	9.11	0.16	0.26	14.01	159.5	
6	Calibration	2	C:\Users\rin0	1000_100-150.1	9.11	0.16	0.26	14.01	159.5	
7	Calibration	2	C:\Users\rin0	1000_100-150.2	9.11	0.16	0.26	14.01	159.5	
8	Calibration	2	C:\Users\rin0	1000_100-150.3	9.11	0.16	0.26	14.01	159.5	
9	Calibration	3	C:\Users\rin0	1005_010-020.0	8.56	0.077	0.56	2.56	58.7	
10	Calibration	3	C:\Users\rin0	1005_010-020.1	8.56	0.077	0.56	2.56	58.7	
11	Calibration	3	C:\Users\rin0	1005_010-020.2	8.56	0.077	0.56	2.56	58.7	
12	Calibration	3	C:\Users\rin0	1005_010-020.3	8.56	0.077	0.56	2.56	58.7	
13	Calibration	4	C:\Users\rin0	1005_050-070.0	8.78	0.077	0.31	1.97	269.2	
14	Calibration	4	C:\Users\rin0	1005_050-070.1	8.78	0.077	0.31	1.97	269.2	
15	Calibration	4	C:\Users\rin0	1005_050-070.2	8.78	0.077	0.31	1.97	269.2	
16	Calibration	4	C:\Users\rin0	1005_050-070.3	8.78	0.077	0.31	1.97	269.2	
17	Calibration	5	C:\Users\rin0	1005_070-090.0	8.78	0.071	0.19	3.21	170.2	
18	Calibration	5	C:\Users\rin0	1005_070-090.1	8.78	0.071	0.19	3.21	170.2	
19	Calibration	5	C:\Users\rin0	1005_070-090.2	8.78	0.071	0.19	3.21	170.2	
20	Calibration	5	C:\Users\rin0	1005_070-090.3	8.78	0.071	0.19	3.21	170.2	
21	Calibration	6	C:\Users\rin0	1010_020-030.0	7.01	0.019	0.3	2.56	30.4	▼



Load Method Components Spectra **Parameters** Validate Graph Report Store Method Optimize Settings

Preprocessing in individual regions (PS)

Set

Preprocessing in calibration regions

**No spectral data preprocessing**

**Mean Centering**

Calibration regions

	from	to	Spacing
1	<b>4000</b>	<b>600</b>	1
2			1

Interactive Region Selection

Clear Selected Regions

View spectra

Display Preprocessed Spectra

Show all spectra

Sample Statistics

PCA





Load Method Components Spectra Parameters **Validate** Graph Report Store Method Optimize Settings

Validation parameters

	Component	Max. Rank	Use
1	pH_H2O	20	<input checked="" type="checkbox"/>
2	EC	20	<input checked="" type="checkbox"/>
3	OC	20	<input checked="" type="checkbox"/>
4	P_Olsen	20	<input checked="" type="checkbox"/>
5	PBI	20	<input checked="" type="checkbox"/>
6	Exch_Ca	20	<input checked="" type="checkbox"/>
7	Exch_Mg	20	<input checked="" type="checkbox"/>
8	Exch_K	20	<input checked="" type="checkbox"/>

Cross Validation

No. of samples leaving out: 1

Validate

Calculation status





- Load Method
- Components
- Spectra
- Parameters
- Validate
- Graph
- Report
- Store Method
- Optimize
- Settings

Graph page

Marker size:

Method protection

Store spectra in Quant 2 method file

Use this option only if you want to protect a method in the mode 'Enlarge method' or 'Change parameters'.

Select preprocessing options for Optimize

- No spectral data preprocessing
- Constant offset elimination
- Straight line subtraction
- Vector normalization (SNV)
- Min-Max normalization
- Multiplicative scattering correction
- First derivative
- Second derivative
- First derivative + Straight line subtraction
- First derivative + Vector normalization (SNV)

Smoothing points:

Maximum test range:

Run optimization in background

User defined optimization regions

NIR regions (max 5)

A,B regions (max 10)



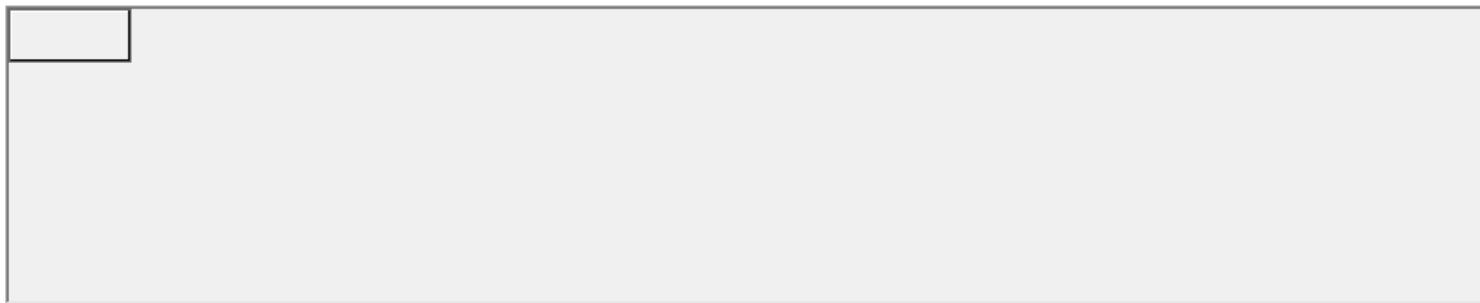
Load Method | Components | Spectra | Parameters | Validate | Graph | Report | **Store Method** | Optimize | Settings

Parameters

**Store only Spectra List + Parameters** ▼

**Store Method**

Store validation results



Spectral residuals

No spectral data preprocessing

Selected datapoints: 1658

Spectra (total): 416

Calibration spectra: 416

Test spectra: 0

	from	to	Spacing
1	4000	600	1

Load Method Components Spectra Parameters Validate Graph Report **Store Method** Optimize Settings

**Store Quant 2 Method**

Look in: **MIR\_Calibration**

Name	Date
Trials	2/03/2019
BaseSat.q2	19/03/2019
Ca_Mg.q2	19/03/2019
Clay.q2	3/03/2019
EC.q2	15/03/2019
ECEC.q2	19/03/2019
ECEC_Clay.q2	19/03/2019
ESP.q2	2/03/2019
Exch_Acid.q2	15/03/2019
Exch_Ca.q2	16/03/2019
Exch_K.q2	16/03/2019
Exch_Mg.q2	19/03/2019
Exch_Na.q2	16/03/2019

File name: **Sample & Component list.Q2**

Files of type: OPUS Quant 2 Methods

Save Cancel



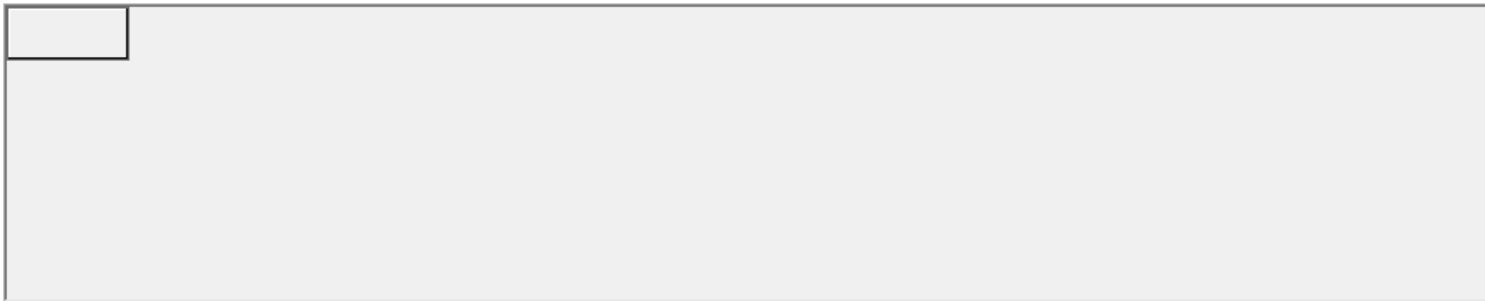
Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

Parameters

Store only Spectra List + Parameters

Store Method

Store validation results



Spectral residuals

No spectral data preprocessing

Selected datapoints: 1658

Spectra (total): 416

Calibration spectra: 416

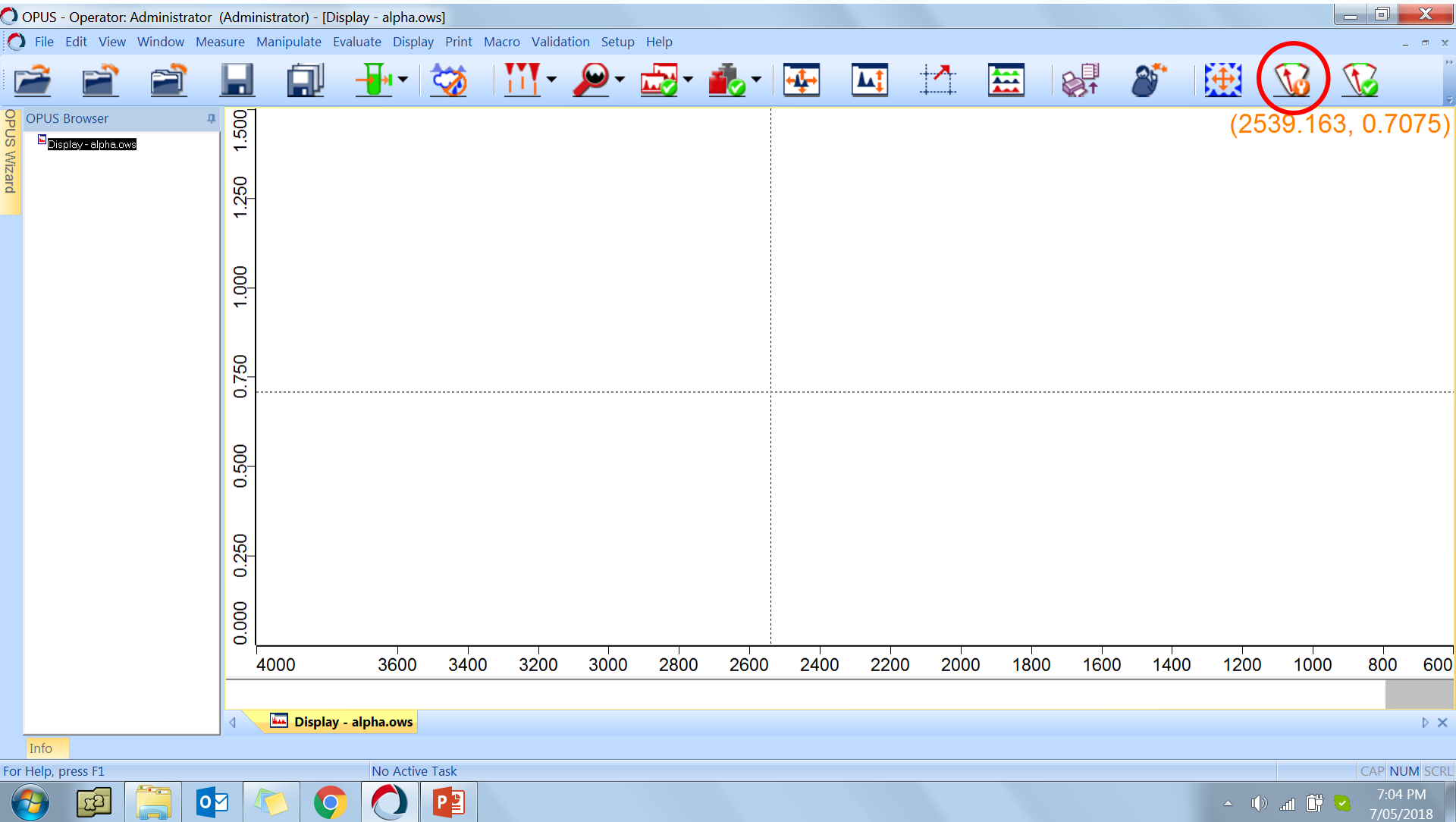
Test spectra: 0

	from	to	Spacing
1	4000	600	1

- This completes setting up the calibration template

# Calibrate each soil property in turn

In this example calibrate pH\_H2O



Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

Load Method

Load existing validation results

General information

Spectra (total): 0

Samples (total): 0

Calibration spectra: 0

Calibration samples: 0

Test spectra: 0

Test samples: 0

Components: 0

Frequency ranges: 0

Selected datapoints: 0

Preprocessing:

No spectral data preprocessing



Look in: MIR\_Calibration

- Recent Places
- Desktop
- Libraries
- Computer
- Network

Name	Date
Exch_Acid.q2	15/0
Exch_Ca.q2	16/0
Exch_K.q2	16/0
Exch_Mg.q2	19/0
Exch_Na.q2	16/0
Exch_Sum.q2	16/0
OC.q2	16/0
P_Olsen.q2	16/0
PBI.q2	16/0
pH_H2O.q2	1/0:
<b>Sample &amp; Component list.q2</b>	15/0
Sand.q2	3/0:
Silt.q2	3/0:

Preview:

QUANT	HISTORY
-------	---------

File name: Sample & Component list.q2

Files of type: OPUS Quant 2 Methods

Open

Cancel

Date of Measurement

Filename: Sample & Component list.q2



Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

Load Method

Load existing validation results

#### General information

Spectra (total): 416

Samples (total): 104

Calibration spectra: 416

Calibration samples: 104

Test spectra: 0

Test samples: 0



Components: 21

Frequency ranges: 1

Selected datapoints: 1658

Preprocessing:

No spectral data preprocessing



Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

Add Spectra

Change Path

Copy Spectra

Window

Set Sample Numbers

Set Data Set

Comp. Correlations

Print

	Data Set	Sample	Path	File Name	pH_H2O	EC	OC	P_Olsen	PBI	E
1	Calibratio	1	C:\Users\rin0	1000_000-	7.85	0.224	1.85	52.28	93	9.4
2	Calibratio	1	C:\Users\rin0	1000_000-	7.85	0.224	1.85	52.28	93	9.4
3	Calibratio	1	C:\Users\rin0	1000_000-	7.85	0.224	1.85	52.28	93	9.4
4	Calibratio	1	C:\Users\rin0	1000_000-	7.85	0.224	1.85	52.28	93	9.4
5	Calibratio	2	C:\Users\rin0	1000_100-	9.11	0.16	0.26	14.01	159.5	6.8
6	Calibratio	2	C:\Users\rin0	1000_100-	9.11	0.16	0.26	14.01	159.5	6.8
7	Calibratio	2	C:\Users\rin0	1000_100-	9.11	0.16	0.26	14.01	159.5	6.8
8	Calibratio	2	C:\Users\rin0	1000_100-	9.11	0.16	0.26	14.01	159.5	6.8
9	Calibratio	3	C:\Users\rin0	1005_010-	8.56	0.077	0.56	2.56	58.7	14
10	Calibratio	3	C:\Users\rin0	1005_010-	8.56	0.077	0.56	2.56	58.7	14
11	Calibratio	3	C:\Users\rin0	1005_010-	8.56	0.077	0.56	2.56	58.7	14
12	Calibratio	3	C:\Users\rin0	1005_010-	8.56	0.077	0.56	2.56	58.7	14
13	Calibratio	4	C:\Users\rin0	1005_050-	8.78	0.077	0.31	1.97	269.2	12
14	Calibratio	4	C:\Users\rin0	1005_050-	8.78	0.077	0.31	1.97	269.2	12
15	Calibratio	4	C:\Users\rin0	1005_050-	8.78	0.077	0.31	1.97	269.2	12
16	Calibratio	4	C:\Users\rin0	1005_050-	8.78	0.077	0.31	1.97	269.2	12
17	Calibratio	5	C:\Users\rin0	1005_070-	8.78	0.071	0.19	3.21	170.2	15
18	Calibratio	5	C:\Users\rin0	1005_070-	8.78	0.071	0.19	3.21	170.2	15
19	Calibratio	5	C:\Users\rin0	1005_070-	8.78	0.071	0.19	3.21	170.2	15
20	Calibratio	5	C:\Users\rin0	1005_070-	8.78	0.071	0.19	3.21	170.2	15
21	Calibratio	6	C:\Users\rin0	1010_020-	7.01	0.019	0.3	2.56	30.4	3.2



# Set Data Set

First test sample:

Block length (test samples):

Gap (calibration samples):

Leave 'Exclude' spectra

Set Test Samples

Clear Test Spectra

Exchange: Test <-> Calibration

## Automatic selection of test samples

Test samples (in %):

Select Test Samples

## Set selected spectra on:

Calibration

Set Data Set

## Set color on page Graph for selected spectra

Blue



Color

Set Color

Clear Color Settings

## Reference values from spectrum file

Get Reference Values

Special Setting...

Exit



### Set Spectra on 'Excluded'



All spectra which have no defined component value (blank) or -1 or 0 as value for the selected component, are set on 'Excluded'.

1.

Blank

2.

pH\_H2O

3.

Set

4.

Exit

Optimize Settings

Copy Spectra

Window

Comp. Correlations

Print

OC	P_Olsen	PBI	E
1.85	52.28	93	9.4
1.85	52.28	93	9.4
1.85	52.28	93	9.4
1.85	52.28	93	9.4
0.26	14.01	159.5	6.8
0.26	14.01	159.5	6.8
0.26	14.01	159.5	6.8
0.26	14.01	159.5	6.8
0.26	14.01	159.5	6.8
0.56	2.56	58.7	14
0.56	2.56	58.7	14
0.56	2.56	58.7	14
0.56	2.56	58.7	14
0.56	2.56	58.7	14
0.56	2.56	58.7	14
0.56	2.56	58.7	14
0.56	2.56	58.7	14
0.56	2.56	58.7	14

12	Calibratio	3	C:\Users\rin0	1005_010-	8.56	0.077	0.56	2.56	58.7	14
13	Calibratio	4	C:\Users\rin0	1005_050-	8.78	0.077	0.31	1.97	269.2	12
14	Calibratio	4	C:\Users\rin0	1005_050-	8.78	0.077	0.31	1.97	269.2	12
15	Calibratio	4	C:\Users\rin0	1005_050-	8.78	0.077	0.31	1.97	269.2	12
16	Calibratio	4	C:\Users\rin0	1005_050-	8.78	0.077	0.31	1.97	269.2	12
17	Calibratio	5	C:\Users\rin0	1005_070-	8.78	0.071	0.19	3.21	170.2	15
18	Calibratio	5	C:\Users\rin0	1005_070-	8.78	0.071	0.19	3.21	170.2	15
19	Calibratio	5	C:\Users\rin0	1005_070-	8.78	0.071	0.19	3.21	170.2	15
20	Calibratio	5	C:\Users\rin0	1005_070-	8.78	0.071	0.19	3.21	170.2	15
21	Calibratio	6	C:\Users\rin0	1010_020-	7.01	0.019	0.3	2.56	30.4	3.5



# Set Data Set

First test sample:

Block length (test samples):

Gap (calibration samples):

Leave 'Exclude' spectra

Clear Test Spectra

Exchange: Test <-> Calibration

Set Test Samples

## Automatic selection of test samples

Test samples (in %):

Select Test Samples

## Set selected spectra on:

Calibration

Set Data Set

## Set color on page Graph for selected spectra

Blue



Color

Set Color

Clear Color Settings

## Reference values from spectrum file

Get Reference Values

Special Setting...

Exit



Load Method Components Spectra Parameters **1. Validate** Graph Report Store Method Optimize Settings

## Validation parameters

	Component	Max. Rank	Use
1	pH_H2O	20	<input checked="" type="checkbox"/>
2	EC	20	<input type="checkbox"/>
3	OC	20	<input type="checkbox"/>
4	P_Olsen	20	<input type="checkbox"/>
5	PBI	20	<input type="checkbox"/>
6	Exch_Ca	20	<input type="checkbox"/>
7	Exch_Mg	20	<input type="checkbox"/>
8	Exch_K	20	<input type="checkbox"/>

2.

Cross Validation

No. of samples leaving out: 1

3. **Validate**

## Calculation status



Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

Validation parameters

	Component	Max. Rank	Use
1	pH_H2O	20	<input checked="" type="checkbox"/>
2	EC	20	<input type="checkbox"/>
3	OC		
4	P_Ols		
5	PBI		
6	Exch		
7	Exch		
8	Exch		

Cross Validation

No. of samples leaving out: 1

Set Validation Name

Please enter a name for the validation.

1. pH\_H2O\_O

2. OK

Cancel

Validation names

- Hint: Use parameter name (pH\_H2O) and "O" (for original)





Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

pH\_H2O\_0

Validation

Calibration

Prediction / True

pH\_H2O

Rank: 11

Rec: 11

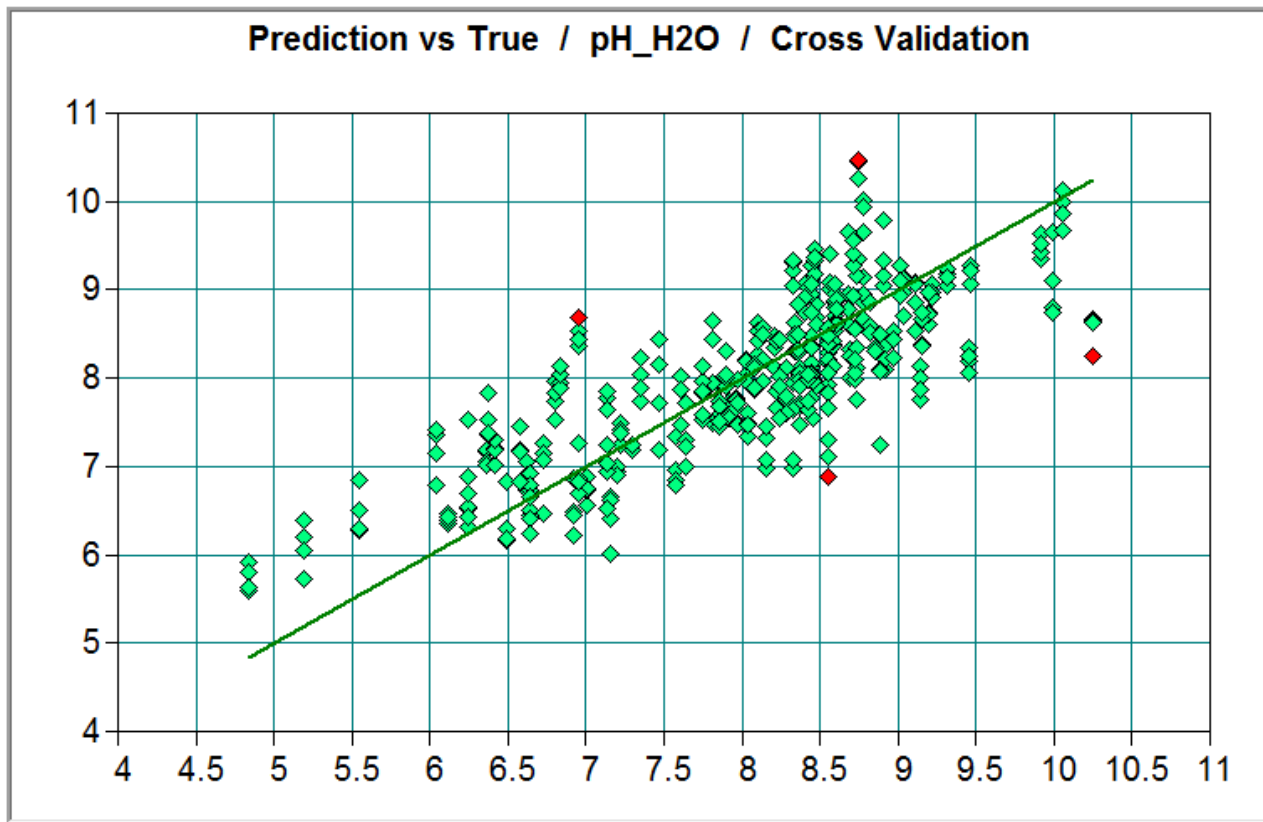
R<sup>2</sup>:  
63.48

RMSECV:  
0.638

RPD:  
1.65

Bias:  
0.00663

Line  
 Color



Window

Print

Save

Spectra

Loadings

Special...

- Next optimize the calibration



Load Method | Components | Spectra | Parameters | Validate | Graph | Report | Store Method | Optimize | Settings

Use Parameters

pH\_H2O

1. General A

2. Optimize

Num...	RMSE...	Rank	Regions	Preprocessing
151	0.425	8	4000.9 - 3659.3 3320.6 - 1275.5 93...	First derivative + Vector normali...
148	0.425	8	4000.9 - 1275.5 936.8 - 595.2	First derivative + Vector normali...
161	0.425	8	4000.9 - 3659.3 3320.6 - 2979.1 26...	First derivative + Vector normali...
153	0.425	8	4000.9 - 2979.1 2638.9 - 1275.5 93...	First derivative + Vector normali...
102	0.425	9	4000.9 - 2979.1 2638.9 - 1275.5 93...	First derivative
143	0.425	9	4000.9 - 2979.1 2638.9 - 595.2	First derivative + Vector normali...
110	0.425	9	4000.9 - 3319.2 2638.9 - 1275.5 93...	First derivative
139	0.426	9	4000.9 - 595.2	First derivative + Vector normali...
141	0.426	9	4000.9 - 3659.3 3320.6 - 595.2	First derivative + Vector normali...
109	0.426	9	4000.9 - 3659.3 3320.6 - 2979.1 26...	First derivative
152	0.426	8	4000.9 - 3319.2 2980.5 - 1275.5 93...	First derivative + Vector normali...
160	0.426	8	4000.9 - 3659.3 2980.5 - 1275.5 93...	First derivative + Vector normali...
97	0.426	9	4000.9 - 1275.5 936.8 - 595.2	First derivative
101	0.426	9	4000.9 - 3319.2 2980.5 - 1275.5 93...	First derivative
142	0.426	9	4000.9 - 3319.2 2980.5 - 595.2	First derivative + Vector normali...
100	0.427	9	4000.9 - 3659.3 3320.6 - 1275.5 93...	First derivative
150	0.427	8	3660.8 - 1275.5 936.8 - 595.2	First derivative + Vector normali...
159	0.427	8	3320.6 - 1275.5 936.8 - 595.2	First derivative + Vector normali...

Optimize status



- Now wait a long time (sometimes hours)



Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

2. Use Parameters

pH\_H2O

General A

Optimize

Optimization number

1.

Num...	RMSE...	Rank	Regions	Preprocessing
95	0.594	17	2976 - 2294.1 1616.4 - 1273.4	First derivative
90	0.599	18	3657.9 - 3314.9 2976 - 2294.1 1616...	First derivative
86	0.611	17	3996.7 - 3314.9 2976 - 2294.1 1616...	First derivative
91	0.613	18	3996.7 - 3655.8 2976 - 2294.1 1616...	First derivative
84	0.617	16	3657.9 - 2294.1 1616.4 - 1273.4	First derivative
92	0.619	16	3996.7 - 3314.9 2637.1 - 2294.1 16...	First derivative
83	0.62	18	3996.7 - 2294.1 1616.4 - 1273.4	First derivative
14	0.634	11	3996.7 - 3314.9 2976 - 1953.2 1616...	No spectral data preprocessing
15	0.636	11	3996.7 - 2973.9 2637.1 - 1953.2 16...	No spectral data preprocessing
36	0.637	9	3996.7 - 1953.2 1616.4 - 593.6	Vector normalization (SNV)
8	0.637	10	3996.7 - 1953.2 1616.4 - 593.6	No spectral data preprocessing
1	0.638	11	3996.7 - 593.6	No spectral data preprocessing
22	0.639	13	3996.7 - 3655.8 2976 - 1953.2 1616...	No spectral data preprocessing
85	0.64	15	3996.7 - 3655.8 3316.9 - 2294.1 16...	First derivative
19	0.64	10	3996.7 - 1953.2 1616.4 - 1273.4 93...	No spectral data preprocessing
3	0.64	12	3996.7 - 3655.8 3316.9 - 593.6	No spectral data preprocessing
23	0.642	10	3996.7 - 3314.9 2637.1 - 1953.2 16...	No spectral data preprocessing
27	0.643	10	3996.7 - 3314.9 2976 - 1953.2 1616...	No spectral data preprocessing

▪ Select top result – OPUS thinks this is the best one

Optimize status

Optimization finished

Load Method Components Spectra Parameters **Validate** Graph Report Store Method Optimize Settings

## Validation parameters

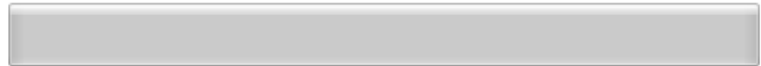
	Component	Max. Rank	Use
1	pH_H2O	20	<input checked="" type="checkbox"/>
2	EC	20	<input type="checkbox"/>
3	OC	20	<input type="checkbox"/>
4	P_Olsen	20	<input type="checkbox"/>
5	PBI	20	<input type="checkbox"/>
6	Exch_Ca	20	<input type="checkbox"/>
7	Exch_Mg	20	<input type="checkbox"/>
8	Exch_K	20	<input type="checkbox"/>

Cross Validation

No. of samples leaving out: 1

**Validate**

## Calculation status





Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

Validation parameters

	Component	Max. Rank	Use
1	pH_H2O	20	<input checked="" type="checkbox"/>
2	EC	20	<input type="checkbox"/>
3	OC		
4	P_Ols		
5	PBI		
6	Exch		
7	Exch		
8	Exch		

Cross Validation

No. of samples leaving out: 1

Set Validation Name



Please enter a name for the validation.

1. pH\_H2O\_A95

2. OK

Cancel

Validation names

- Hint: Use parameter name and "A95" (95 was optimization number)



Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

pH\_H2O\_A95

Validation

Calibration

Prediction / True

pH\_H2O

Rank: 17

Rec: 17

R<sup>2</sup>: 68.3

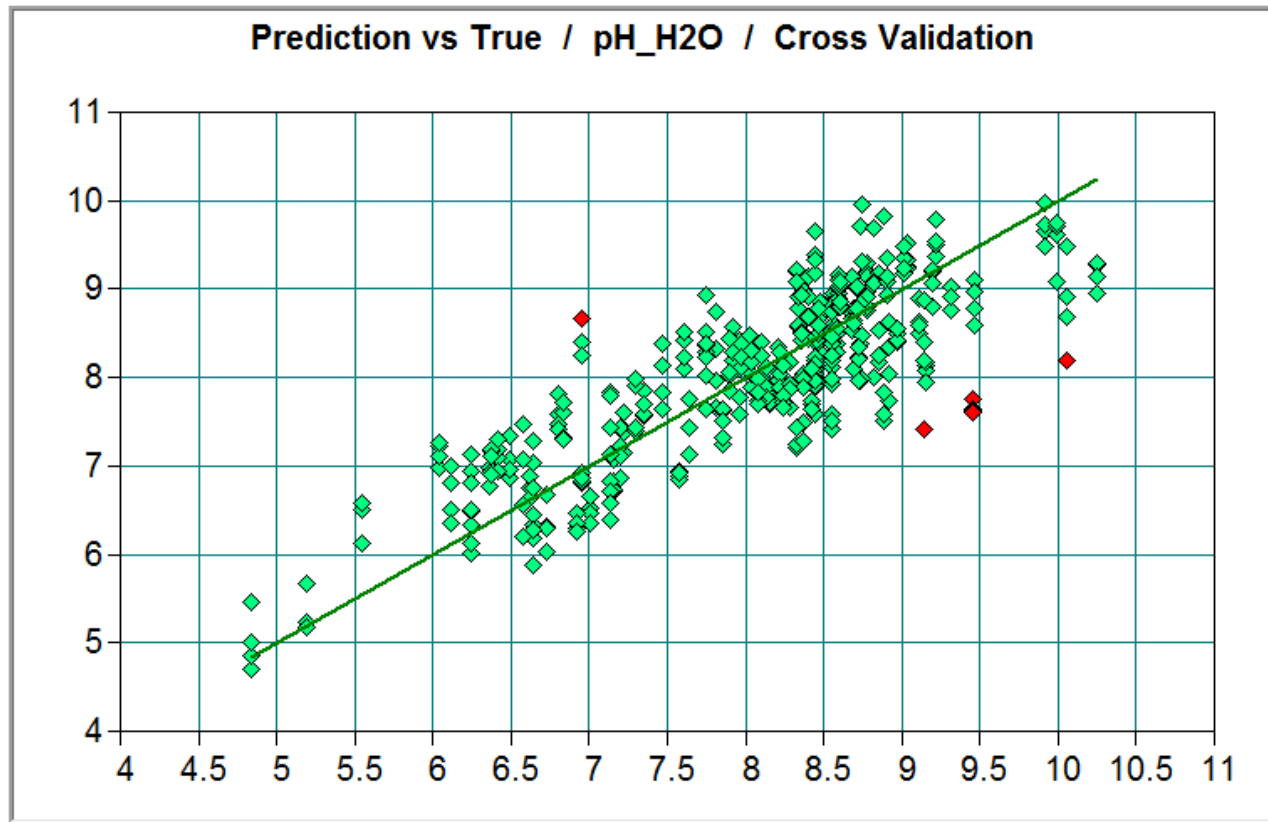
RMSECV: 0.594

RPD: 1.78

Bias: 0.0112

Line

Color



Window

Print

Save

Spectra

Loadings

Special...



Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

2. Use Parameters

pH\_H2O

General A

Optimize

Optimization number

Num...	RMSE...	Rank	Regions	Preprocessing
95	0.594	17	2976 - 2294.1 1616.4 - 1273.4	First derivative
90	0.599	18	3657.9 - 3314.9 2976 - 2294.1 1616...	First derivative
86	0.611	17	3996.7 - 3314.9 2976 - 2294.1 1616...	First derivative
91	0.613	18	3996.7 - 3655.8 2976 - 2294.1 1616...	First derivative
1. 84	0.617	16	3657.9 - 2294.1 1616.4 - 1273.4	First derivative
92	0.619	16	3996.7 - 3314.9 2637.1 - 2294.1 16...	First derivative
83	0.62	18	3996.7 - 2294.1 1616.4 - 1273.4	First derivative
14	0.634	11	3996.7 - 3314.9 2976 - 1953.2 1616...	No spectral data preprocessing
15	0.636	11	3996.7 - 2973.9 2637.1 - 1953.2 16...	No spectral data preprocessing
36	0.637	9	3996.7 - 1953.2 1616.4 - 593.6	Vector normalization (SNV)
8	0.637	10	3996.7 - 1953.2 1616.4 - 593.6	No spectral data preprocessing
1	0.638	11	3996.7 - 593.6	No spectral data preprocessing
22	0.639	13	3996.7 - 3655.8 2976 - 1953.2 1616...	No spectral data preprocessing
85	0.64	15	3996.7 - 3655.8 3316.9 - 2294.1 16...	First derivative
19	0.64	10	3996.7 - 1953.2 1616.4 - 1273.4 93...	No spectral data preprocessing
3	0.64	12	3996.7 - 3655.8 3316.9 - 593.6	No spectral data preprocessing
23	0.642	10	3996.7 - 3314.9 2637.1 - 1953.2 16...	No spectral data preprocessing
27	0.643	10	3996.7 - 3314.9 2976 - 1953.2 1616...	No spectral data preprocessing

Find first optimization that has a lower rank

▪ Select top result – OPUS thinks this is the best one

Optimize status

Optimization finished

- Validate first optimization that has a lower rank
- Examine  $R^2$ . If it is less than 1% lower than the top optimization:
  - Consider using it instead of the top optimization

- Next Select <General B> on the Optimize sheet
- Wait while the optimization runs



- When finished, select top optimization and validate
- Then select first optimization that has a lower rank and validate
- On the Graph sheet, compare  $R^2$  and rank of all the selected optimizations and choose which to use as a calibration



Load Method | Components | Spectra | Parameters | Validate | Graph | Report | Store Method | Optimize | Settings

**pH\_H2O\_B360**

Validation  
 Calibration

Compare different optimizations using this pull-down menu

Prediction / True

pH\_H2O

Rank: **8**

Rec: 8

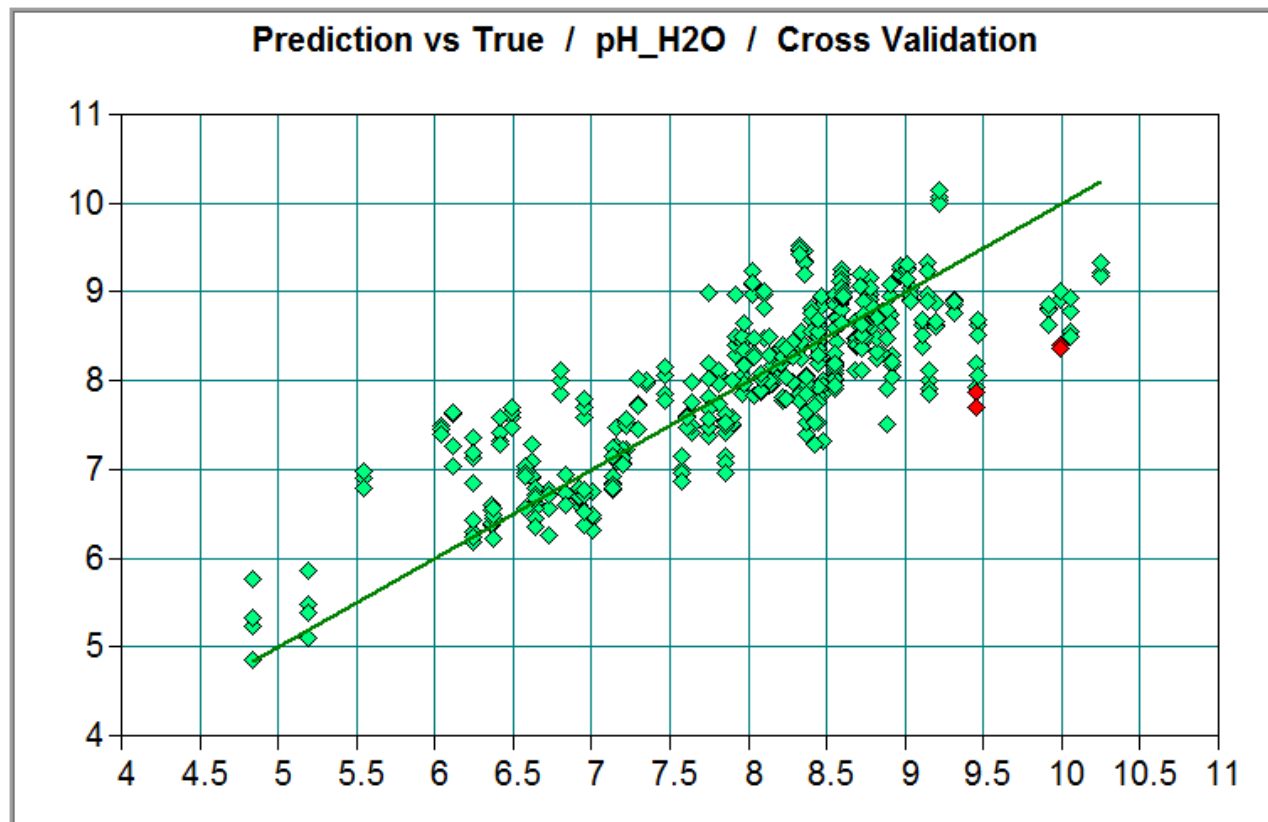
**R<sup>2</sup>: 65.98**

RMSECV: 0.616

RPD: 1.71

Bias: 0.000274

- Line
- Color



Window

Print

Save

Spectra

Loadings

Special...



Load Method Components Spectra Parameters Validate Graph Report **Store Method** Optimize Settings

Parameters for Prediction

Select chosen optimization here

1. pH\_H2O\_A95

Cross Validation

2. **Store Method**

Store validation results

	Component	Rec. Rank	Rank (Method)	R <sup>2</sup>	RMSECV	MD Limit	Use
1	pH_H2O	17	17	68.3	0.594	0.16	<input checked="" type="checkbox"/>
2	EC						<input type="checkbox"/>
3	OC						<input type="checkbox"/>
4	P_Olsen						<input type="checkbox"/>
5	PBI						<input type="checkbox"/>
6	Exch Ca						<input type="checkbox"/>

Spectral residuals

First derivative (17 Pts.)

Selected datapoints: 501

Spectra (total): 416

Calibration spectra: 416

Test spectra: 0

	from	to	Spacing
1	2976	2294.1	1
2	1616.4	1273.4	1

Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

### Select Validation Results

The results of the selected validations will be stored.

1. **Select All**

pH\_H2O\_O  
pH\_H2O\_A95  
pH\_H2O\_A84  
pH\_H2O\_B303  
pH\_H2O\_B360

2. **OK**

Cancel

Select all optimizations here for record-keeping: only the one selected previously will be used

	C
1	pl
2	E
3	O
4	P
5	P
6	E

Spectral r

Sele

Cal

Spacing
1
1

Load Method Components Spectra Parameters Validate Graph Report Store Method Optimize Settings

### Store Quant 2 Method

Look in: MIR\_Calibration

Name	Date
Exch_Acid.q2	15/0
Exch_Ca.q2	16/0
Exch_K.q2	16/0
Exch_Mg.q2	19/0
Exch_Na.q2	16/0
Exch_Sum.q2	16/0
OC.q2	16/0
P_Olsen.q2	16/0
PBI.q2	16/0
pH_H2O.q2	1/03
Sample & Component list.q2	15/0
Sand.q2	3/02
Silt.q2	3/02

File name: 1. pH\_H2O.q2 2. Save

Files of type: OPUS Quant 2 Methods

Cancel

- The chosen calibration is now saved as a “Method”, and can be used to predict the pH of other samples based on their MIR spectra

# Thank you

**CSIRO Agriculture and Food**

Anthony Ringrose-Voase

Principal Research Consultant

t +61 2 6246 5956

e [anthony.ringrose-voase@csiro.au](mailto:anthony.ringrose-voase@csiro.au)

CSIRO Agriculture and Food

[www.csiro.au](http://www.csiro.au)

