

Technical assessment of the eReefs biogeochemical (BGC) simulation [gbr4_H2p0_B3p1_Chyd_Dcrt] against observations and comparison with BGC version B3p0

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1. Summary:

This document is a technical assessment of eReef model version: gbr4_H2p0_B3p1_Chyd_Dcrt. [GBR4: model grid with approximate 4 km grid resolution, H2p0: hydrodynamic model version 2.0, B3p1: biogeochemical model version 3.0, Chyd: Department of Environment and Science QLD catchment model, 2011, Catchment version is May 2019 catchment, Dhnd: deployment in hindcast mode].

The eReefs model configuration and results used in this assessment is available via https://research.csiro.au/ereefs/models/ where the assessment link is labelled "Reanalysis [GBR4_H2p0_B3p1_Chyd_Dhnd]"

The model simulation period is from 1 Dec 2010 to November 2019 In this technical assessment it is compared with observations and the previous biogeochemical model version (gbr4_H2p0_B3p0_Chyd_Dcrt)

Access to the assessment is from the links in the table of contents.

Model metrics updated 23 April 2020 Word document updated: Thursday, 27 August 2020



2. Abbreviations

AIMS	Australian Institute of Marine Science
B2p0	B2p0: biogeochemical model version 2.0
B3p1	B3p1: biogeochemical model version 3.0
CDOM	colour dissolved organic matter
Chl a	chlorophyll a
CTD	Conductivity Temperature Depth profiler
d2	Statistical metric, aka Willmott index (see page Error! Bookmark not defined.)
DIN	dissolved inorganic nitrogen
DIN	Dissolved inorganic nitrogen (NH3 plus NOx)
DIP	dissolved inorganic phosphorus
DOC	dissolved organic carbon
DON	dissolved organic nitrogen
DOP	dissolved organic phosphorus
GBR	Great Barrier Reef
gbr4_H2p0_B3p1_Cb	gbr4 : model grid with approximate 4 km grid resolution, H2p0: hydrodynamic
	model version 2.0, B3p1: biogeochemical model version 3.1, Cb: catchment
	model baseline version using empirical SOURCE Catchments (2019 version)
IMOS	Integrated Marine and Observing System
Kd(PAR)	light attenuation coefficient
LTM	AIMS long term monitoring site
mae	mean absolute error
mape	mean absolute percentage error
MMP	AIMS Marine Monitoring Program
NH3	ammonia
NOx	nitrate plus nitrite
NRS	IMOS National reference station within the model grid these are Yongala
	(GBRYON) and North Stradbroke Island (GBRNSI)
NSI	North Stradbroke Island
NTU	Nephelometric Turbidity Unit
PON	particulate organic nitrogen
POP	particulate organic phosphorus
QA/QC	quality assurance/quality control
rms	root mean square
secchi	measurement of water transparency (depth in m)
TSS	total suspended solids
Willmott	statistical metric





3. River and catchments in eReefs reanalysis 2020

• Rivers and catchment model with hydro flow catchment loads

• Extra rivers in B3p1 where catchment is included as point source loads

• Rivers in hydrodynamic model, some without flow, no catchment model data

Figure 1 Map of Queensland rivers included in eReef model reanalysis version B3p1. Includes extra rivers for B3p1 in light blue



4. AIMS Long Term Monitoring Marine Monitoring Program 2020 sites (pink), 2019/2020 sites (black) and IMOS NRS stations (NRSYON, NRSNSI)





eReefs BGC model B3p1 2020





eReefs BGC model B3p1 2020



5. eReefs biogeochemical model schematic



Figure 2. The eReefs modelling system, showing the linkages between hydrodynamic, wave, sediment and the optical and biogeochemical models, as well as the individual linkages within the biogeochemical model. The optically-active components are identified with orange colouring (adapted from Baird et al., 2016b).

6. Model skill metrics used in reanalysis

To evaluate model skill, we consider; bias, the root mean square (RMS) error, the mean absolute error (MAE). and the modified Willmott index or 'd2' (Willmott et al., 1985). The Willmott index uses the sum of absolute values.

Model bias assesses whether the simulated variables are under- or over-predicting observed values. The RMS error is a measure of the absolute magnitude of the "error"/square deviation averaged over the time-series. An RMS or MAE of 0 indicates a perfect fit.

The Willmott index of agreement is designed to quantify errors that are unevenly distributed in time or space and reduce the influence of errors during periods of large observed mean or variance. The Willmott index is the ratio of the mean absolute error and the mean absolute deviation about the observed mean and varies between 0 and 1. A value of 1 indicates a perfect match (x = y), and 0 indicates no agreement.

Willmott = 1 - [$\sum |x - y|$)/ [$\sum |x - \bar{y}|$) + (|y - $\bar{y}|$)]

where x and y are vectors or arrays of time series data (x =observed, y = modelled).

A Willmott index above 0.7 is regularly obtained for high resolution models with high spatial and temporal observations for physical parameters such as salinity and temperature. In most cases for the eReefs model the salinity and temperature index was \geq 0.8 when compared with observations (Appendix 1 of Herzfeld et al., 2016).









Figure 3 Metrics for Long Term Monitoring Sites simulated Chlorophyll a against observations



























8. Simulated Secchi depth assessment against AIMS Long Term Monitoring (includes scatter plots)























Figure 5 Scatter plot of observed Secchi for Long Term Monitoring sites and NRS sites (Yongala and North Stradbroke) assessment against simulated Secchi for model version 3p1



9. Simulated DIP assessment against AIMS Long Term Monitoring





Figure 6 Metrics for Long Term Monitoring Sites simulated DIP against observations


























10. Simulated NOx assessment against AIMS Long Term Monitoring





Figure 7 Metrics for Long Term Monitoring sites simulated NO3 against observations

























11. Simulated NH4 assessment against AIMS Long Term Monitoring



Figure 8 Metrics for Long Term Monitoring Sites simulated NHx (ammonia + ammonium) against observations

























12. Simulated DON assessment against Long Term Monitoring





Figure 9 Metrics for Long Term Monitoring Sites simulated DON against observations
























13. Simulated DOP assessment against Long Term Monitoring







Figure 10 Metrics for Long Term Monitoring Sites simulated DOP against observations

























Simulated TSS assessment against Long Term Monitoring 14.





Figure 11 Metrics for Long Term Monitoring Sites simulated TSS against observations

Greents1-0m

PortD-0m

Snap-10m

PortD-15m

Fairlead-0m

FitzCoral-0n HighIsl-0m HighIsl-10m HighIsl-20m Russell-10m Russell-20m

Dunklsl-5m

Pelorus-0n

Russell-0m

PineIsI-20m

PineIs1-0m

PelicanIsI-5n










































15. Simulated Chl *a* assessment against National Reference Stations

Figure 12 Metrics for IMOS NRS sites simulated Chlorophyll a against observation

16. Simulated Chl a assessment against National Reference Stations









Figure 13 Metrics for IMOS NRS sites simulated Chlorophyll a against observation







17. Simulated Chl *a* and Fluorescence assessment against AIMS















0.5

0 Dec/10

May/12

Sep/13

Feb/15

Jun/16

Nov/17

Mar/19









Figure 15 Scatter plot of observed Fluorescence for AIMS MMP assessment against simulated Chl a for model version 3p1





Figure 16 Scatter plot of observed Fluorescence for AIMS MMP assessment against simulated Chl a for model version 3p1

18. Simulated Turbidity assessment against AIMS MMP Turbidity

W



Figure 17 Metrics for AIMS MMP sites simulated turbidity against observations













19. Simulated NOx assessment against NRS: Yongala and NSI





run1 mean bias:-0.87 run2 mean bias:-0.78 run1 =/home/mgdata/mgdata/scratch/gbr4_bgc/gbr4_H2p0_B3p0_Chyd_Dcrt/final/ run2 =/bowen/eReefsWQ/scratch/gbr4_H2p0_B3p1_Cq3b/





Figure 18 Metrics for NRS sites simulated NOx against observations







20. Simulated NHx assessment against NRS: Yongala and NSI





run1 mean bias:-0.83 run2 mean bias:-0.54 run1 =/home/mgdata/mgdata/scratch/gbr4_bgc/gbr4_H2p0_B3p0_Chyd_Dcrt/final/





Figure 19 Metrics for NRS sites simulated NHx (ammonia + ammonium) against observations





Simulated DIP assessment against NRS: Yongala and NSI 21.





run1 mean bias:-1.00 run2 mean bias:-0.70 run1 =/home/mgdata/mgdata/scratch/gbr4_bgc/gbr4_H2p0_B3p0_Chyd_Dcrt/final/ run2 =/bowen/eReefsWQ/scratch/gbr4_H2p0_B3p1_Cq3b/







Figure 20 Metrics for NRS sites simulated DIP against observations





22. Simulated alkalinity assessment against NRS: Yongala and NSI





run1 mean bias:-3.38 run2 mean bias:-5.48 run1 =/home/mgdata/mgdata/scratch/gbr4_bgc/gbr4_H2p0_B3p0_Chyd_Dcrt/final/ run2 =/bowen/eReefsWQ/scratch/gbr4_H2p0_B3p1_Cq3b/





Figure 21 Metrics for NRS sites simulated alkalinity against observations







23. Parameter values for gbr4_H2p0_B3p1_Chyd_Dcrt

The following tables list the parameter values used in the gbr4_H2p0_B3p1_Chyd_Dcrt biogeochemical model simulation



Parameter description	Symbol	Units	Value	Reference
Phytoplankton	_			
Chl-specific scattering coefficient. for microalgae	bphy	m ⁻¹ (mg Chl a m ⁻³) ⁻¹	0.2	Typical microalgae value, Kirk (1994)
Natural (linear) mortality rate, large phytoplankton	PhyL mL	d ⁻¹	0.1	Not attributed
Natural (linear) mortality rate in sediment, large	PhyL mL sed	d-1	10	Not attributed
phytoplankton	<i>,</i>			
Natural (linear) mortality rate, small phytoplankton	PhyS_mL	d-1	0.1	Not attributed
Natural (linear) mortality rate in sediment, small	PhyS_mL_sed	d-1	1	Not attributed
phytoplankton				
Respiration as a fraction of umax	Plank_resp	none	0.025	Not attributed
Radius of the large phytoplankton cells	PLrad	m	0.000004	Not attributed
Maximum growth rate of PL at Tref	PLumax	d-1	1.4	CSIRO Parameter Library
Ratio of xanthophyll to chl a of PL	PLxan2chl	mg mg ⁻¹	0.81	CSIRO Parameter Library
Radius of the small phytoplankton cells	PSrad	m	0.000001	Not attributed
Maximum growth rate of PS at Tref	PSumax	d-1	1.6	CSIRO Parameter Library
Ratio of xanthophyll to chl a of PS	PSxan2chl	mg mg ⁻¹	0.51	CSIRO Parameter Library
Trichodesmium				
DIN conc below which Trichodesmium N fixes	DINcrit	mg N m ⁻³	10	Lower end of Robson et al., (2013) 4-20 mg N m ⁻³
Maximum density of Trichodesmium	p_max	kg m ⁻³	1050	Not attributed
Minimum density of Trichodesmium	p_min	kg m ⁻³	900	Not attributed
Radius of Trichodesmium colonies	Tricho_colrad	m	0.000005	Not attributed
Critical Trichodesmium above which quadratic	Tricho_crit	mg N m ⁻³	0.0002	Not used in code
mortality applies				
Linear mortality for Trichodesmium in sediment	Tricho_mL	d-1	0.1	Not attributed
Quadratic mortality for Trichodesmium due to	Tricho_mQ	d⁻¹ (mg N m⁻³)⁻¹	0.1	At steady-state, indep. of temp, Tricho_N ~ Tricho_umax / Tricho_mQ = 0.27 /
phages in water column				0.405 = 0.7 mg N m ⁻³ ~ 0.1 mg Chl m ⁻³
Trichodesmium grazing preference	Tricho_pref	none	0	Not attributed
Radius of Trichodesmium colonies	Tricho_rad	m	0.000005	Not attributed
Sherwood number for the Trichodesmium	Tricho_Sh	none	1	Not attributed
dimensionless				
Maximum growth rate of Trichodesmium at Tref	Tricho_umax	d-1	0.2	Robson et al., 2013 + Parameter library
Ratio of xanthophyll to chl a of Trichodesmium	Trichoxan2chl	mg mg ⁻¹	0.5	Subramaniam et al. 1999. LO 44:618-627
Microphytobenthos				
Respiration as a fraction of umax	Benth_resp	none	0.025	Not attributed
Radius of the MPB cells	MBrad	m	0.00001	Not attributed
Maximum growth rate of MB at Tref	MBumax	d-1	0.839	CSIRO Parameter Library
Ratio of xanthophyll to chl a of MPB	MBxan2chl	mg mg ⁻¹	0.81	Not attributed
Natural (quadratic) mortality rate,	MPB_mQ	d ⁻¹ (mg N m ⁻³) ⁻¹	0.0001	SS argument
microphytobenthos, applied in sediment				


Parameter description	Symbol	Units	Value	Reference
Zooplankton				
Growth efficiency, large zooplankton	ZL_E	none	0.426	CSIRO Parameter Library, [0.341 (0.017900) Baird and Suthers, 2007 from Hansen et al (1997) LO 42: 687-704]
Fraction of growth inefficiency lost to detritus, large zooplankton	ZL FDG	none	0.5	Not attributed
Fraction of mortality lost to detritus, large zooplankton	ZL FDM	none	1	Not attributed
Natural (guadratic) mortality rate, large zooplankton	ZL mQ	d ⁻¹ (mg N m ⁻³) ⁻¹	0.012	Not attributed
Diel vertical migration rate of ZL	ZLdvmrate	m d ⁻¹	0	Not attributed
Grazing technique of large zooplankton	ZLmeth	none	rect	Not attributed
Light at which the	ZLpar	mol photons m ⁻² s ⁻¹	1.00E-12	Not attributed
Radius of the large zooplankton cells	ZLrad	m	0.00032	Not attributed
Swimming velocity for large zooplankton	ZLswim	m s⁻¹	0.003	Not attributed
Maximum growth rate of ZL at Tref	ZLumax	d-1	1.33	Not attributed
Growth efficiency, small zooplankton	ZS_E	none	0.462	CSIRO Parameter Library [0.3080000 (0.026600) Baird and Suthers,
	_			2007 from Hansen et al (1997) LO 42: 687-704]
Fraction of growth inefficiency lost to detritus, small zooplankton	ZS_FDG	none	0.5	Not attributed
Fraction of mortality lost to detritus, small zooplankton	ZS FDM	none	1	Not attributed
Natural (quadratic) mortality rate, small zooplankton	ZS_mQ	d ⁻¹ (mg N m ⁻³) ⁻¹	0.02	Not attributed
Grazing technique of small zooplankton	ZSmeth	none	rect	Not attributed
Radius of the small zooplankton cells	ZSrad	m	0.000005	Not attributed
Swimming velocity for small zooplankton	ZSswim	m s⁻¹	0.0002	Not attributed
Maximum growth rate of ZS at Tref	ZSumax	d-1	4	Not attributed
Coral				
Quadratic mortality rate of coral polyp	CHmort	(g N m ⁻³) ⁻¹ d ⁻¹	0.01	Not attributed
Nitrogen-specific area of coral polyp density	CHpolypden	m2 g N ⁻¹	2	Not attributed
Fraction of Host death translocated.	CHremin	-	0.5	Not attributed
Max. growth rate of Coral at Tref	CHumax	d-1	0.05	Not attributed
Linear mortality rate of Zooxanthellae	CSmort	d-1	0.04	Not attributed
Radius of the Zooxanthellae	CSrad	m	0.000005	Not attributed
Fraction of Zooxanthellae growth to Host.	CStoCHfrac	-	0.9	Gustafsson et al. (2013) Ecol. Mod. 250: 183-194
Max. growth rate of Zooxanthellae at Tref	CSumax	d-1	0.4	Not attributed
Maximum daytime net coral calcification	k_day_coral	mmol C m ⁻² s ⁻¹	0.0132	Anthony et al. (2013), Biogeosciences 10:4897-4909, Fig 5A: 50, 50, 35 55 mmol m ⁻² h ⁻¹ for Acropora aspera n=4
Grid scale to reef scale ratio	CHarea	m m ⁻¹	0.1	Not attributed
Maximum night time net coral calcification	k_night_coral	mmol C m ⁻² s ⁻¹	0.0069	Anthony et al. (2013), Biogeosciences 10:4897-4909, Fig 5A: 20, 30,
Data coofficient for plantton untake by corols	Colonk		2	20, 30 mmol m ⁻² h ⁻¹ for Acropora aspera n=4 Bibos (2002), DADAMETER, library analysis/Bibos and Atkinson (2007)
Rate coefficient for plankton uptake by corais	Splank	m a -	3	Coral Reefs 26: 413-421
Macroalgae				
Maximum growth rate of MA at Tref	MAumax	d ⁻¹	1	Not attributed
Natural (linear) mortality rate, macroalgae	MA_mL	d ⁻¹	0.01	Not attributed
Nitrogen-specific area of macroalgae leaf	MAleafden	m ² g N ⁻¹	1	Not attributed



Parameter description	Symbol	Units	Value	Reference
Seagrass				
Half-saturation of SG N uptake in SED	SG KN	mg N m ⁻³	420	Lee and Dunton (1999) 1204-1215. Table 3 Zostera
Half-saturation of SG P uptake in SED	SG KP	mg P m ⁻³	96	Gras et al. (2003) Aquatic Botany 76:299 ⁻³ 15. Thalassia testudinum.
Natural (linear) mortality rate, seagrass	SG mL	d ⁻¹	0.03	Fourguean et al. (2003) Chem. Ecol. 19: 373-390. Thalassia leaves with one component decay
Critical shear stress for SG loss	SG tau critical	N m^{-2}	1	NESP project
Time-scale for critical shear stress for SG loss	SG tau efold	S	43200	NESP project
Half-saturation of SGD N uptake in SED	SGD KN	mg N m ⁻³	420	Not attributed
Half-saturation of SGD P uptake in SED	SGD_KP	mg P m ⁻³	96	Not attributed
Natural (linear) mortality rate, aboveground SGD	SGD mL	d-1	0.06	NESP project
Critical shear stress for SGD loss	SGD tau critical	N m ⁻²	1	NESP project
Time-scale for critical shear stress for SGD loss	SGD_tau_efold	S	43200	NESP project
Fraction (target) of SGD biomass below-ground	SGDfrac	-	0.25	Duarte (1999) Aquatic Biol. 65: 159-174, Halophila ovalis.
Nitrogen-specific leaf area of SGD	SGDleafden	m² g N⁻¹	1.9	Halophila ovalis: leaf dimensions from Vermaat et al. (1995)
Compensation irradiance for Halophila	SGDmlr	mol m ⁻²	1.5	NESP project
Sine of nadir Deep Seagrass canopy bending angle	SGDorient	-	1	No source
Natural (linear) mortality rate, belowground SGD	SGDROOT_mL	d-1	0.004	NESP project
Maximum depth for Halophila roots	SGDrootdepth	m	-0.05	NESP project
Halophila seed biomass as fraction of 63 % cover	SGDseedfrac	-	0.01	Not attributed
Time scale for seagrass translocation	SGDtransrate	d-1	0.0333	Loosely based on Zostera marine Kaldy et al., 2013 MEPS 487:27-39
Maximum growth rate of SGD at Tref	SGDumax	d-1	0.4	x2 nighttime, x2 for roots.
Fraction (target) of SG biomass below-ground	SGfrac	-	0.75	Babcock (2015) Zostera capricornii
Half-saturation of SGH N uptake in SED	SGH_KN	mg N m ⁻³	420	Not attributed
Half-saturation of SGH P uptake in SED	SGH_KP	mg P m ⁻³	96	Not attributed
Natural (linear) mortality rate, seagrassH	SGH_mL	d ⁻¹	0.06	Fourquean et al. (2003) Chem. Ecol. 19: 373 ⁻³ 90. Thalassia leaves with one component decay
Critical shear stress for SGH loss	SGH_tau_critical	N m ⁻²	1	NESP project
Time-scale for critical shear stress for SGH loss	SGH_tau_efold	S	43200	NESP project
Fraction (target) of SGH biomass below-ground	SGHfrac	-	0.5	Babcock 2015, Halophila ovalis
Nitrogen-specific area of seagrass leaf	SGHleafden	m2 g N ⁻¹	1.9	Halophila ovalis: leaf dimensions from Vermaat et al. (1995)
Compensation irradiance for SG	SGHmlr	mol m ⁻²	2	Not attributed
Sine of nadir Halophila canopy bending angle	SGHorient	-	1	No source
Natural (linear) mortality rate, seagrassH	SGHROOT_mL	d-1	0.004	Fourquean et al. (2003) Chem. Ecol. 19: 373-390. Thalassia roots with one component decay
Maximum depth for Halophila roots	SGHrootdepth	m	-0.08	Roberts (1993) Aust. J. Mar. Fresh. Res. 44:85-100.
Halophila seed biomass as fraction of 63 % cover	SGHseedfrac	-	0.01	Not attributed
Time scale for seagrass translocation	SGHtransrate	d-1	0.0333	Loosely based on Zostera marine Kaldy et al., 2013 MEPS 487:27-39
Maximum growth rate of SGH at Tref	SGHumax	d-1	0.4	x2 night-time, x2 for roots.
Nitrogen-specific area of seagrass leaf	SGleafden	m2 g N⁻¹	1.5	Zostera capricornia: leaf dimensions Kemp et al (1987) Mar Ecol. Prog. Ser. 41:79-86.
Compensation irradiance for SG	SGmlr	mol m ⁻²	4.5	Not attributed
SGorient	SGorient		0.5	Not attributed
Natural (linear) mortality rate, seagrass	SGROOT_mL	d-1	0.004	Fourquean et al. (2003) Chem. Ecol. 19: 373-390. Thalassia roots with one component decay
Maximum depth for Zostera roots	SGrootdepth	m	-0.15	Roberts (1993) Aust. J. Mar. Fresh. Res. 44:85-100.
Seagrass seed biomass as fraction of 63 % cover	SGseedfrac	-	0.01	No source
Time scale for seagrass translocation	SGtransrate	d-1	0.0333	Loosely based on Zostera marine Kaldy et al., 2013 MEPS 487:27-39
Maximum growth rate of SG at Tref	SGumax	d-1	0.4	x2 nighttime, x2 for roots.



Parameter description

Symbol

Units

Value Reference

Tref	Deg C	20	CSIRO Parameter Library
Q10	none	2	CSIRO Parameter Library
TKEeps	m² s-3	0.000001	Not attributed
xco2_in_air_dum	ppmv	396.48	Mean 2013 at Mauna Loa: htttrp://co2now.org/current-co2/co2-now/
Light_lambda	nm	Various*	Approx. 20 nm resolution with 10 nm about 440 nm. PAR (400-700) is integral of bands 2-22 (290 310 330 350 370 390 410 430 440 450 470 490 510 530 550 570 590 610 630 650 670 690 710 800)*
NtoCHL	g N (g Chl a)-1	7	Represents a C:Chl ratio of 39.25, Baird et al. (2013) Limnol. Oceanogr. 58: 1215-1226.
N2	mg N m⁻³	2000	Robson et al. (2013)
F_LD_RD	none	0.19	Not attributed
F_LD_DOM	none	0.1	Not attributed
F_RD_DOM	none	0.05	Not attributed
r_DetPL	d-1	0.04	Not attributed
r DetBL	d-1	0.001	Not attributed
r_RD	d-1	0.001	Not attributed
r_DOM	d-1	0.0001	Achieves approx. SS of global ocean at 20 C.
KO_aer	mg O m⁻³	256	Not attributed
r_nit_wc	d-1	0.1	Not attributed
r_nit_sed	d-1	20	Not attributed
KO_nit	mg O m⁻³	500	Not attributed
Pads_r	d-1	0.04	Not attributed
Pads_Kwc	mg P kg TSS ⁻¹	30	Not attributed
Pads_Ksed	mg P kg TSS ⁻¹	74	Not attributed
Pads_KO	mg O m⁻³	2000	Not attributed
Pads_exp	none	1	Not attributed
r_den	d-1	0.8	Not attributed
KO_den	mg O m⁻³	10000	Not attributed
r_immob_PIP	d-1	0.0012	Not attributed
EpiDiffCoeff	m² s ⁻¹	3.00E-07	Not attributed
EpiDiffDz	m	0.0065	Not attributed
ageing_decay	d-1	1	Not attributed
anti_ageing_decay	d-1	0.1	Not attributed
dissCaCO3_sed	mmol C m ⁻² s ⁻¹	0.001	Anthony et al. (2013), Biogeosciences 10:4897-4909, Fig 5E: -1 2 3 6 mmol m $^{ m 2}$ h $^{ m 1}$
acdom443star	m ² mg C ⁻¹	0.00013	Not attributed
C2Chlmin	wt/wt	20	Not attributed
SWRscale	none	1	Not attributed
ROSthreshold	-	5.00E-04	Not attributed
r_RD_NtoP	-	2	Not attributed
r_DOM_NtoP	-	1.5	Not attributed
	Tref Q10 TKEeps xco2_in_air_dum Light_lambda NtoCHL N2 F_LD_RD F_LD_DOM F_RD_DOM r_DetPL r_DetBL r_RD r_DOM KO_aer r_nit_wc r_nit_sed KO_ater r_nit_wc r_nit_sed KO_ater r_nit_wc r_nit_sed KO_ater r_ates KO_den r_den KO_den r_immob_PIP EpiDiffCoeff EpiDiffDz ageing_decay anti_ageing_decay anti_ageing_decay dissCaCO3_sed acdom443star C2Chlmin SWRscale ROSthreshold r_RD_NtoP r_DOM_NtoP	TrefDeg CQ10noneTKEeps $m^2 s^{-3}$ $xco2_in_air_dum$ ppmvLight_lambdanmNtoCHLg N (g Chl a) ⁻¹ N2mg N m ⁻³ F_LD_RDnoneF_LD_DOMnoneF_RD_DOMnoner_DetPLd ⁻¹ r_DetBLd ⁻¹ r_TOMd ⁻¹ r_Sedd ⁻¹ r_MCd ⁻¹ r_ADd ⁻¹ r_DetBLd ⁻¹ r_ADd ⁻¹ r_DetBLd ⁻¹ r_ADd ⁻¹ r_Boysmg O m ⁻³ r_nit_wcd ⁻¹ Pads_rd ⁻¹ Pads_Komg P kg TSS ⁻¹ Pads_Komg O m ⁻³ Pads_Komg O m ⁻³ r_dend ⁻¹ KO_denmg O m ⁻³ r_idend ⁻¹ KO_denmg O m ⁻³ r_idendm ¹ koodenmg O m ⁻³ r_idendd ⁻¹ koodenmg O m ⁻³ r_idendd ⁻¹ Kozelemoner_idendmati_ageing_decayd ¹ d ¹ koodenmorekoodenmol C m ⁻² s ⁻¹ action443starm ² mg C ¹ c2chlminwt/wtsWRscalenoneROSthreshold-r	TrefDeg C20Q10none2TKEeps $m^2 s^{-3}$ 0.000001 $xco2_in_air_dum$ ppmv396.48light_lambdanmVarious*NtoCHLg N (g Chl a)-17N2mg N m ⁻³ 2000F_LD_RDnone0.19F_LD_DOMnone0.05r_DetPLd ⁻¹ 0.001r_RDd ¹ 0.001r_DetBLd ⁻¹ 0.001r_DoMd ¹ 0.001r_DetBLd ⁻¹ 0.001r_DoMd ¹ 0.001r_DoMd ¹ 0.001r_Sedd ⁻¹ 0.1r_nit_wcd ⁻¹ 0.1r_nit_sedd ⁻¹ 0.04Pads_rmg O m ⁻³ 500Pads_rd ⁻¹ 0.04Pads_Kwcmg P kg TSS ⁻¹ 30Pads_KoDmg O m ⁻³ 2000Pads_expnone1r_dend ⁻¹ 0.8KO_denmg O m ⁻³ 10000r_imob_PIPd ⁻¹ 0.0012EpiDiffCoeffm ² s ⁻¹ 3.00E-07EpiDiffDzm0.0065ageing_decayd ⁻¹ 1acti_ageing_decayd ⁻¹ 0.0013C2Chlminwt/wt20SWRscalenone1ROSthreshold-5.00E-04r_RD_NtoP-2r_DOM_NtoP-1.5